A Complete Bibliography of Publications in the

Nelson H. F. Beebe
University of Utah
Department of Mathematics, 110 LCB
155 S 1400 E RM 233
Salt Lake City, UT 84112-0090
USA
Tel: +1 801 581 5254
FAX: +1 801 581 4148

E-mail: beebe@math.utah.edu, beebe@acm.org, beebe@computer.org (Internet)
WWW URL: http://www.math.utah.edu/~beebe/

08 October 2019
Version 1.02

Title word cross-reference

1 [ARV+18, FFG+18, SPE+17a, VMR+19]. 2 [MF16b]. 3
[CCQ+18, CBM+16, LE16, MPMP16, MWSM18, MWSM19, MPW+19,
RGOS+16, SZL+16, Sør17, SBP+16, WZG+17, WWT18]. Cde55 [JRH+16].
[LCD+17]. Zyg11 [BHS+16, Bra16]. 1 [CNC+18]. 2
[CCCL17, CWZ+15, Dic17, GPD+19, HQW15, JJW17, KML+15, LLW+15,
MBC+19, DR19, SKZ+18a, Yu19]. 2a [XJG+17]. 3 [RSC+19]. α
[BNS+17, BKG5+15, BAGM17, CST+16, CHI+15, CKS+15, DR16,
FBBRCA+18, FRP+17, GSP+18, HDA+17, JCK+19, K15a, KT15b,
LLC+17, LBV+17, MB17a, MSS+17, NNK+15, PAC+15, Qi17, RKK+18,
STR+18, SSV+18, SFZ+17, TTU+17, WWZ+18, WIS+17, YTGA16, ZT15].
\( \beta \) [AGB+19, ACG+17, ARV+18, BAGM17, CHS+17, CSG+15, CIK+17,
DVS+17, DKA+16, FWL+17, FVF+16, GBD+18, HAK+15, JLB+18,
JJB+19, Les15u, LJ17b, LDR+19, LSS+15, LLC+17, MA17, MSS+17,
PhHS+16, PTK16, PLH18, PW19, PAC+15, RR+17, Sho15s, SPJ+15,
SHVO+18, SLG+18, TSK+18, TSK+19, VXF+15, WWZ+18, WEQ+15,
WGHE+18, XWZ+15, XMJ+19, XTT+18, YYY+15, ZT15]. \( \beta_{1-42} \) [QYC+17].
\( \delta \) [DVS+17, GSD+15, RKK+18, WDM+15]. \( \gamma \) [CKS+15, IZZ+18, cLNF+16,
LDR+19, LBV+17, MSL+17, MSL16, SVD+15, Sho15-68, SKZ+18b]. \( \kappa \)
[Hu15, LAMACE+17, MCS+15, YGMR+17, ZLG+15, dVGO+16]. \( \mu \)
[NEW+17].

-1 [RKK+18]. -actin [MSS+17, SVD+15, Sho15-68, SPJ+15]. -actinin-
[KT15a, KT15b]. -appendage [FRP+17]. -arrestin [HDA+17, PhHS+16].
-barrel [JLB+18, WEQ+15]. -binding [RSC+19]. -Catenin
[CH+15, WGHE+18, WIS+17, GBD+18, LJ17b, RR+17, MB17a].
-dependent [KT15a, KT15b, CBM+16]. -directed [GSD+15].
-Glycan-dependent [LGH+18]. -induced [VXF+15, XWZ+15]. -Integrins
[JCK+19]. -kinase [GWZ+19, JJB+19]. -like [DVS+17]. -phosphatase
[DZB+18, NMN+15]. -phosphate [HHM15, Les15o, LJS+16a, LJS+16b].
-PIX [LS+15]. -QC [MPA+16]. -Secretase [IZZ+18, LDR+19].
-secretases [CKS+15]. -stimulated [MWSM18, MWSM19]. -synuclein
[CST+16, DR16]. -TAT1 [FBBRCA+18]. -tubulin
[BNS+17, cLNF+16, MSL16, SKZ+18b]. -TuSC [cLNF+16].

/H [MPMP16].

1 [AZS+15, BKH+15, CWL+17, CRPSC+19, CPBG19, CST+16, CYL+18,
CLBB15, CBB15, DDAR+16, DR16, DKA+16, DLBMA+15, FSB+15,
GCJ+15, HBS+15, HAR+15, JNW15, KCB+16, LAMACE+17, MSK+18,
MFP17, MDC+16, NEW+17, NNK+15, OOT+18, QCC+19, SCNTC+18,
Sed15x, Sho15-48, SENL+15, SLH17, TNP+15, TCD+15, XMJ+19, YHG+17,
YKKB17]. 1/APP1 [LRM+19]. 1/Bora [TNP+15]. 1/BUB
[KMLG+15, KMLG+16]. 1/p38 [DKA+16]. 10 [LWH+18]. 103/ [PPK+16].
11 [CNN+17, Les17]. 14-3-3 [BBS+17, Das17, iNLM+19]. 16 [WZR19]. 17
[Juh16, MLM16]. 170 [JNW15]. 1A [BRY+19, KCB+16]. 1b
[IYP+18, PLD+15, HBDW+15]. 1E [THA+16].

2 [BBSA+16, BVR+17, CLV17, CLO+19, DMS+15, DKA+16, EMRS+18, EPF16,
FVF+16, GCC+18, KMK+17a, KMK+17b, NWD+19, SAF+19, WWZ+17].
2/SEDB1 [DMG+19]. 200 [HBWY18]. 203 [LCM+16].

3 [BVR+17, DDAR+16, HBDW+15, KSM+18, LRBB15, Les15t, LCZ+16,


5 [RHC+16, LWH+18, SNGO16]. 5-bisphosphate [GCJ+15]. 5'-Inositol [RHC+16]. 51 [OMKM16]. 53BP1 [LDU+16, BCMM+19, Can19, LCD+17, MAK+16]. 5P [JJW17].

6 [ABPS17, CIS+17, CYL+18, dVGO+16]. 60S [BMW+18]. 65 [DMG+19]. 6a [CYT+18]. 6B [SXT16].

7 [CSC+15, Sho15-34]. 7/ [ZLG+15]. 70/110 [PXN18].

8 [GLSS+15b, GLSS+15a, KKP+17, SKZ+18a]. 84 [SJ16].

actin-based [HH16]. actin-binding [GDB+15, OLT+19].
actin-cytoskeletal [RSC+15]. actin-dependent [DQB+16, YEM+19].
actin-filled [FLLM17]. actin-mediated [MBS+18, vGWC+18]. actinin
[KT15a, KT15b]. action [KHS+16, MSvO17, Sho16-37, Sho18e]. actions
[MSE+17, vHGd+15]. activate [LMPG+15, PYO+18]. activated
[CSS+19, GSP+18, GSM+15, NWFY15, WHS+19, WIS+17, YWW17, YNN18].
activates
[BNB+15, CZZ+15, IBG+15, LM15, MSK+18, MCL+15, TJF18, WFOA15].
Activating [PHKY17, SENL+15, YHG+17]. activation
[AGGSF+16, APS+17, AIS+18, ANM+19, BLG+15, Bob17, BLZ+15,
CWL+17, CPCr+15, CLBB15, DPS+18, DCO+12, DCO+16, FTAB+15,
GBD+18, GLC+19, GAS+15, HMM+19, HB16, HLEM+18, IKT+18, KSL+17,
LH15, LS18, LWH+18, LBV+17, MMW+19, MAK+16, MCS+15, MCGC+15,
MJSB16, NKP+15, PLS+15, PTMP+15, SV16, SSRG18, SK16b, SQC+16,
SS18, TNP+15, TF16, TGQ+17, UFT+15, VXF+15, WG16, WXC+18,
WWZ+18, WWZ+17, Woo18, WKW+15, YTL15, dIRHM+18]. activator
[BC19, GSKL+18, GTD+18, ODH19, RYS+15]. Active
[HLST19, SHVO+18, CL19, GBB+19, KMK+17a, KMK+17b, cLNf+16,
MWT+16, NHA+19, SES+19, THM+19]. Activity
[GSs+17, AFT+19, BMS+17, CIK+17, CYL+18, DKM+15, ESS+17,
FKL+18a, FKL+18b, GHD+17, GBD+18, GLJ+17, GLS+15, HZL+15,
HCN+15, IKK+18, KBB+15, KBB+16, LJ17a, LRBB15, Log17, LWF+15,
MKs+18, MGT+19, MpDN+17, MCL+15, RGM+16, SSH+15, TBJ+17,
TF19, VMR+19, WDM+15, ZAT+19, ZAT+17]. Activity-dependent
[GSs+17, HZL+15]. actomyosin
[CMMB+15, HLHF15, MHY+16, NLBA+15, OKN+16, XS16, Mar16b]. acts
[CSc+15, FG15, HB18, LWH+18, SBC+16a, SBC+16b]. acute
[CHZ+17, GD16, NS15, NKX+15]. ADAM10 [DCO+12, DCO+16].
ADAM10/Kuzbanian [DCO+12, DCO+16]. adapt [PXN18]. adaptability
[Sho17]. adapter [WV18b]. adapters [BhiHS+17]. adaptor [DKR+19a,
DKR+19b, MYT+16, NEW+17, SV16, SD19, VMR+19, WHS+19, ZY16].
adaptors [BDK+18]. adapts [OII18b]. added [Sho15-30]. Adding [Sho15c].
addition [CG17, MOM+18]. adducts [ABGG16]. Adenomatous [JBE+17].
adenosine [Xjg+17]. adenylyl [CS16a]. Adherens [SOII18, BPH+18,
CAP+16, ES18, GPA1A+18, KLS+19, TE15, TCD+15, WW16]. Adhesion
[Bea16, KG15, AMS+17, BP19c, CTI+19, CLBB15, DBC+15, DCM+17, ES18,
EVK+19, HfwV18, HHS+16, IKA+15, JAH118, JBE+17, JIB+19, KSG+16,
KS17, KOV+16a, KOV+16b, LLK+17, LDM17, LCM+16, LM19, LBJ+19,
MCO+19, POE+16, PMG+17, RbZ18, SSPD15, WCl+18, WIS+17, ZBl8].
adhesion-based [JKA+15]. adhesions [FBPN+18, FKG+19, GGF+19,
Sho15i, Sho16s, SHVO+18, SZR+15, TLMG+15]. adhesive [HVH+19].
adipocyte [SQB+15]. adipocytes [BBC+16]. adipogenesis [EW17].

avoid [ML15b, RM16, Sho16z]. GTD \textit{baton} [O’D18d].
dVGO RGOS [BCG barcoding] [Bra16, Rab17, VHB18].
RGM [DATI18, FLLM17, HH16, ISK+15, KDR+19, KSM+17, RGOS+16, THG19].
axons [GTW+15, VPD+16, ZHP+19].

autophagy-deficient [WCY+16a, WCY+16b].
autophagy-dependent [KSG+16].
autophagy-related [CD18].
Auwerx [Inf18c].
availability [AB18, PKKB17].
available [Av18a].
avenues [MG18].
avoid [ML15b, RM16, Sho16z].
avoiding [JW19].
award [Sho16a].
avay [Bao-Liang]

autophagy-dependent [LAMACE+17].
B1 [BHS+16, LTL+18, SHG+18, dVGO+16].
B2 [LTC+18].
B3 [KBK19, LIW+19].
B55 [MBG+18b, CHB+16].
B56 [HBM+19].
BACE1 [LDR+19].
Back [TL17, XS16, Jor16g, JGCAC+15, Sho15x].
back-to-back [XS16].
bacteria [BLL15, CSM17].
bacterial

BHK17, ISK+15, TLMG+15, VQ17, ZWB+19].
Badovinac [Pow15j].

BAF [KL19].
BAG3 [ALLY+17].
BAIAP3 [Ser17, ZM+17].
BAK [CLV17, iHM+17].
balance

DCB+15, EMRS+18, Mes16, Sed15t, Sho16l, SAO+17].
Balanced [Les15b].
balances [MTG18].
Balancing [Will15, TF19].
Bal巴拉 [O’D18c].
ballet

[Hen19].
bang [FLG+18, TNL18, Jan18].
Bao [O’D18a].

BAR [O’D18a].

KJON+17, SE19, SZK+19, UMC+15, UMC+17, WYV+19, WMB+15].
barcoding [BCG+19].
Bar [Jor16e].
barrel [JLB+18, WEQ+15].
Barres [AD18].
barrier [HSK+19, JKD+19, KHRL17, KSM+18, KBB+15, KBB+16, MRGB+16, NIN+19, Sho15-40, SSE18, SCK+19, SLN+15, TCD+15].
barrier-to-autointegration [HSK+19].

Basal

[LMTZ+18, MGT+19, BG3+16, GBRH15, HTK+16, PVP18, RDH+19].
based [DATI18, FLLM17, HH16, ISK+15, KJA+15, KDR+19, KSM+17, RGOS+16, THG19].
Basement [CC19, CPB+16, JCK+19].
basic [SHo16c].
basis [ATRG19, GFvA+15, MKA+17].

Baton [O’D18d].
Bayonets [ZB19].
BBF2H7 [ITN+17].

BBSome

[YNN18].
Bcl [CLV17].
Bcl-2 [CLV17].

BDNF
[BLZ⁺¹⁵, FTAB⁺¹⁵, ODH19]. BDNF-signaling [ODH19]. be
[Jor16b, LPWK15]. BEACH [LLW⁺¹⁷]. BEACH-containing [LLW⁺¹⁷].
beacon [PH16]. Bear [Cas16b]. Beata [Inf18a]. beating [BGJ⁺¹⁶].
becomes [Ser17]. before [LMC⁺¹⁸, SSRG18]. Beginning [Ger15]. behave
[Les15b]. behavior [CRZ⁺¹⁶, IBG⁺¹⁵, Wil15]. behind [OM19]. Ben
[AD18]. bending [TBJ⁺¹⁷]. bent [MOM⁺¹⁸]. best [NA16]. bet
[DAG⁺¹⁵, vHG⁺¹⁵]. Bet1 [MHA⁺¹⁹]. BethAnn [IO18]. better
[Les15b, NA16]. Between [Lov18, ABF⁺¹⁶, BFPD19, BDLB15, CANG⁺¹⁷,
CKS⁺¹⁵, CCH⁺¹⁷, GSRG⁺¹⁸, HGL⁺¹⁷, Inf18a, KTK⁺¹⁸, MGO⁺¹⁸,
PMRM17, PUY⁺¹⁹, SLW⁺¹⁸, SCG17, SZR⁺¹⁵, TE15, UDΗ⁺¹⁶, VGB⁺¹⁷].
Beyond [CD18, CC19, DR16, Gar15a]. Bhalla [Sed16d]. bias
Bidirectional [BMF⁺¹⁸, RFG⁺¹⁹]. bifurcated [CKKG17]. big
[FB15, FLG⁺¹⁸, FA16, MCI15, Jan18, TNK18]. BIK [CHL⁺¹⁹]. Bin1
[NiYT⁺¹⁶]. binders [HCML15]. Binding [MCL⁺¹⁵, BHB⁺¹⁸, BDLB15,
BPW15, Bob17, BS17b, CCQ⁺¹⁸, CBH⁺¹⁵, FCB⁺⁰⁹, FCB⁺¹⁹, GHF⁺¹⁶,
GDB⁺¹⁵, GLC⁺¹⁹, HKM⁺¹⁵, KGN⁺¹⁵, KDV⁺¹⁵, MDOS19, OLT⁺¹⁹,
PKC⁺¹⁶, QCC⁺¹⁹, QZY⁺¹⁹, RSC⁺¹⁹, SER⁺¹⁵, SMK⁺¹⁸, SG17, SIYM⁺¹⁸,
WV18b, HSN⁺¹⁶, LHY⁺¹⁹, NiG⁺¹⁸, NP0⁺¹⁷, YLND⁺¹⁶]. binds
[BBSA⁺¹⁶, CPEE⁺¹⁵, FLG⁺¹⁸, GDB⁺¹⁵, HBDW⁺¹⁵, MAJ⁺¹⁷, YVM18,
ZWS⁺¹⁶, vBMG⁺¹⁵]. bioavailability [SAF⁺¹⁹]. biochemical [ECAB⁺¹⁶].
bioenergetics [BBW⁺¹⁸, QJP⁺¹⁷]. biogenesis [BBJ⁺¹⁸, Bob18, CGD⁺¹⁸,
FWL⁺¹⁷, HSB⁺¹⁹, HAR⁺¹⁵, JLB⁺¹⁸, JHC⁺¹⁶, MGE⁺¹⁵, NP15, Sho16-29,
TTC⁺¹⁶, TF16, VYB⁺¹⁹, VKT⁺¹⁵, WEQ⁺¹⁵, ZNR⁺¹⁸, ZWB⁺¹⁹].
bioenergetics [BBW⁺¹⁸, QJP⁺¹⁷]. biogenesis [BBJ⁺¹⁸, Bob18, CGD⁺¹⁸,
FWL⁺¹⁷, HSB⁺¹⁹, HAR⁺¹⁵, JLB⁺¹⁸, JHC⁺¹⁶, MGE⁺¹⁵, NP15, Sho16-29,
TTC⁺¹⁶, TF16, VYB⁺¹⁹, VKT⁺¹⁵, WEQ⁺¹⁵, ZNR⁺¹⁸, ZWB⁺¹⁹].
biologist [Mar15, O’D17c, She15]. biology [BH15, Cas16a, CZP16, DD18,
Fuc15, Gar15b, GGR15, GD16, HCML15, ISK⁺¹⁵, JDG16, MXS17, May15,
MHW19, O’D17d, O’D18a, O’D19g, RSS15, SSC⁺¹⁹, Sch15, SQ15,
SK16b, SKG17, TGO15, Tar15, TMK18, Tra18, YH15, vS15, Hal15].
Biophysical [HSK⁺¹⁶, ECAB⁺¹⁶]. biorientation
[FTDC17, LJ17a, RGM⁺¹⁶]. biosensor [GPĐ⁺¹⁹, OSL⁺¹⁹]. biosensors
[OSL⁺¹⁹]. biosynthesis [Mes16]. biosynthetic [SJL⁺¹⁹]. bipartite
[SCL⁺¹⁹]. Biphasic [FLN⁺¹⁰, Nig⁺¹⁸, FLN⁺¹⁶]. bipolarity [ZLZD16].
biportion [ATRG19]. Bipotent [TGJ⁺¹⁷]. bisphosphate [GCJ⁺¹⁵].
bistability [DSSF⁺¹⁵]. bite [Sho16a]. black [THG19]. blastocyst [BMC15].
blebbing [HHB17]. blebs [CNS19]. BLM [CNA⁺¹⁷, DKS15, PMHB17].
Blob1 [Tra18]. blobs [NC18]. BLOC [FC16, CMS⁺¹⁵, DDR⁺¹⁶, MFP17].
BLOC-1 [DDR⁺¹⁶, MFP17]. BLOC-2 [CMS⁺¹⁵]. BLOC-3 [DDR⁺¹⁶].
block [XMJ⁺¹⁹]. blocking [VXF⁺¹⁵]. blocks [KKC⁺¹⁹, VLP⁺¹⁵]. Blood
[FG16, NIN⁺¹⁹, SLM⁺¹⁵]. Bloo1 [BMM⁺¹⁹]. blue [BP19a, BP19b].
blueprint [KWB⁺¹⁵]. Blume [Sed15p]. BMP [FVF⁺¹⁶, FG16, VAKB⁺¹⁸].
BMP-2 [FVF⁺¹⁶]. bMunc13 [KMK⁺¹⁷b, KMK⁺¹⁷b]. bMunc13-2
[KMK⁺¹⁷a, KMK⁺¹⁷b]. Bnip3 [GDL⁺¹⁵]. Board [Mar19]. bodies
[BGJ⁺¹⁶, BBK16, HTK⁺¹⁶, MPW⁺¹⁹, SD16a, YIT⁺¹⁵]. body

SXT16, BMC15, CBH+15, DSvNA+15a, DSvNA+15b, GBD+18, GDB+15, HLHFG15, JKD+19, RMS+18, Sho15v, VHB18. Cadherin-6B [SXT16].
cadherin/ [BK+15]. cadherins [KHS+16, SPE+17a]. Caenorhabditis [DRMW17, KFAMR17, LYO15, ZAA17]. Calcium
[ZZ+15, VMP16, BZG+17, CJS+18, GSM+15, KBJ16, MJ16, RYS+15, SD19, SK18b, WHS+19]. calcium-activated [WH19]. calmodulin
[ZZ+15]. calpain [AR+18]. calpain- [AR+18]. Calreticulin [SQB+15]. CaM [SZ+15]. cAMP [CS16a, GCVAGS+18, IdSCB+16, IKK+18]. Can
[Ava18, Bro16, LPWK15, LTC+18, MG16, PCK+17, VHB18, Ver16].
canalizes [DKMV15]. Cancer
[AC+17, BBMM+16, EAW+17, RMTR17, ALY+17, BS18, CC19, CBF+18, DMC+16, DCM+17, GLL+18b, GN18, JPC+17, KKP+17, Les15-30, Lin15, MB15, MCC+15, MSV16, MWSM18, MWSM19, MTC+17, NW+19, O’D18g, PAC+15, QSZ+17a, QSZ+17b, RHC+16, RGOS+16, RMS+18, RRM+17, STR+18, Sch17b, Sed15b, Sed15l, Sho16, TG19, TF19, VWM+18, vV17a].
Cancer-associated [AC+17, BBMM+16, EAW+17]. candle [O’D19c].
candles [O’D19c]. cannibalism [Pas19]. canonical
[DRMW17, DGS+18, HB18]. can’t [Kaw17]. capacity [MPDN+17, PBL+19].
capillary [KJZ+19]. capillary-like [KJZ+19]. capping [AKD+17].
Caprin1 [KPA+16]. capsid [ZZ+18]. capture
[BCH+18, HK15, JIB+19, MAJ+19]. carbon [S16-37]. carboxyl [CAA+17].
carcinogenesis [ZL+15]. carcinoma [LAMACE+17]. cardiact
[ASP+16, BFS+19, CMTH+15, MT19]. Cardiolipin [VGB+17, RXEB+19].
cardiomyocyte [AGGSF+16, DV16]. cardiomyocytes
[ASP+16, DK+16]. cardiomyopathies [MHW19]. cardiomyopathy
[CRC+15]. cardiovascular [CLL+16]. Career [Mar19, O’D17d, Sil16a].
Cargo [KJ17, MFVS18, VKJ+15, CPB19, CBM+16, CCM+19, DMS+15, GM16, ITN+17, KMBO+15, KOK+19, LHT+19, MAJ+17, MGJ+16, MPW+19, SSM+18, SV16, SDHC17, WJ18b].
cargo- [ITN+17].
Cargo-selective [KJ17, VKJ+15]. cargo-sorting [KMBO+15].
cargo-specific [LHT+19]. cargos [YDM+18]. carrier [DBW+17]. carriers
[CPBG17, CC+19, DDAR+16, GKY+17, MICS19, RRH+18]. cartilage
[HPE+19]. cartilage-mediated [HPE+19]. cartography [Tar15].
Cas9 [LYO15, MTN+16]. cascade [CKKG17]. case [Les16h]. caspase
[APS+17, GSP+18, OR17, KKP+17]. caspase-2 [APS+17, OR17].
Caspase-8 [KKP+17]. CAST [HKG+18]. casts [Sho15-33]. catabolism
[SWS+19]. catabolite [ZWW+19]. catalyzed [CR18]. catastrophe
[GCL+15, gXNG+15, gXNG+16]. Catch [Das17]. Catching [SS16, O’D17b].
Catenin [CHI+15, WGHE+18, WIS+17, CSG+15, GBD+18, L17b, RR+17, MB17a, BK+15]. cation [LGY+18]. CatSper [EMB+15].
caught [Sed15c]. cause [BHS+18]. causes
[DSH+18, GBD+18, HKG17, YBZ+18, ZT15, MG18]. causing
[OBS+17, V19]. Caveolae [CMTH+15, JSB+18, Sho18a, TSB+18].
[MOJ16]. Cells
[Sho16d, ALY17, BMM19, BRACA16, Blo19, BPS15, BUPC19, CAKL16, CPCTr15, CNCS18, CSG15, CMTH15, CEM15, Col18, CDF18, DMC16, DSC18, DMH15, DVS17, ESS17, FWL17, FC19, FKL18a, FKL18b, FJ17, GBRH15, GLL18a, GLH18b, GCVAGS18, GN18, GAS18, Haw18, HHT16, HKK19, HKT17, HMC16, IGK16, Inf18b, IBM15, JNW15, KFI8, KKP17, KdBKdK15, KOF16, Les15p, Les15o, Les15r, Les15y, LT18, MTN16, MB17a, MA17, MHA16, MWSM18, MWSM19, MpDN17, MPN18, MT19, Niy16, OSW17, O'D17a, OC15, OPP18, OFP19, Ott16, PW19, PKN15, PHKY17, PMW18, PBS16, PCM16, PMG17, Pow16b, QSZ17a, QSZ17b, RMB18, RZS15, RMS18, RMT17, SD19, STR18, SS16, SZF15, SSH15, Sed15, Sed15a, Sho15-29, Sho15-70, Sho16i, Sho16q, Sho16v, Sho16-27, Sho17g, Sho17k, Sho18d, SRT18, SSE18, SKG16, SKO15].

cells
[ST17, SAK18, SCP17, SHO15-74, SMN16, TGJ17, TCP18, TST17, TBL15, TCWM18, TS15b, TSJ15, TSC15, TALR19, TSK18, TSK19, UD116, VM19, VBJ18a, VB18b, VPD16, VAKB18, VKJ15, VZFG18, WCY16a, WCY16b, WKM15, WHB18, XM19, XTS15, YYM18, YTH17, YLW15, YHS15, ZDM15, ZJM17, ZGS16, ZMC15, ZCH18, dVGO16, vV17a, SW18].

Cellular
[BMS17, Bea16, Blu15b, BOL17, CHL19, FCB19, FCB19, FA16, GF16, HF15, KHS16, KJH18, KBJ16, LZ16, LDR19, Mar17, MR18, MSV17, Pas19, Pow15g, RC15, SBM17, TG15, Tar15].

CENP
[BGH18, KWB15, LRS17, LM16, LBB15, MHD15, NAFM17, WFS15].

[AUTM16, LZ16, Sho16], CM16]. centering
[SXE19, TKM16, ZCH18, AZ19]. centers [JhZbYmP15]. central
[GJW17, LPHH16, WPA18, ZHP19, vBGMG15]. centralspindlin
[ABP19]. centrin [MP17b]. Centrin2 [PM15]. centriole
[AWS18, GJFR16, KMC19, LUC15, LBD18, LTS17, MBG18a, MCL15].

Centrioles [SSR17, BPSK16, Ver16]. Centrobin [OTG18, RGR18].

centromere
[BGH18, VGA15, AFT19, HKT17, KWB15, LBB15, WFS15].

centromere-associated [HKT17]. centromeres
[EKJ16, FFATC15, LM16, Sho15-73, Sho16-36]. centromeric
[NAFM17, Sho15-65]. centrosomal [CANG17].

Centrosome
[DSH18, LMC18, PCP17, Sho17b, BYUJ17, Cas16a, CGY19, LDU16, LJP15, LSJY15, LTS17, MAK16, PSL17, PSP15, RFO16, RMS18, SFZ15, YYM18].

centrosome-unattached [RFO16]. centrosomes
[Les15v, O'D17f, PTMP15, Sed16a, VHB18]. CEP83 [LLY19]. ceramide
[LCTP17]. Cerebellar [DLH19]. cerevisiae [LKM15a, LTRW15, YTL15].

chain
[FML17, HPE19, LDMW15, MFVS18, OKK15, Sho15-37, Sho15-49].

chains [GDV19, LYO15]. chairs [SG17]. challenge [AR15]. challenges

Cliff [Jor16b]. CLIP [JNW15, MRK+18]. CLIP-170 [JNW15]. cloaked [Hyr15]. clock [AWS+18]. closer [Inf18b, MB17a, SZ17a].
closure [DKMV15, Mar16b, TLH+16]. compartments [JBBM16].
coat [Gli17]. coated [GYK+17, MFVS18]. coats [RBP+17]. Cobl [ISL+18].
Cobl-like [ISL+18]. Cocaine [NLH+19]. Cocaine-induced [NLH+19].
cochaperone [ABPS17]. code [DK16, O’D19e]. coding [NPQ+17].
Coenzyme [SLL+19, MM+15]. coexpansion [WF15]. cofactor [PNE+19].
cofilin [ZAT+19, HBDW+15]. Cofilin-dependent [HBDW+15].
cohort [Sho15k]. coincidence [DWH+17a]. Cold [XS18].
Cold-induced [XS18]. Cole [Pow15a]. coli [JBE+17, DBS18]. Collagen [SCL+16, ASM+15, CPB+16, ITN+17, JCK+19, Sho15m, Sho16e, Sho16g].
Collagen-derived [SCL+16]. collateral [MSS+17, RM16]. collectins [JNS+19]. Collective [SM16, DPSG+18, HKH16, MBS+17, PBL+16, SBC+16a, SBC+16b, SMN+16, WCL+18, ZTR+17]. collectively [LM19].
collude [DR19]. colonic [AMT+15]. columnar [LDM+17]. columns [CED+15]. come [Jor16d, O’D19e, Sho16-33]. comes [KBJ16, WS18].
Comestibles [MA17]. Coming [FC16]. command [LZ16, LS16].
compaction [EGY+19, FMS+19, KP18]. compartment [BFS+19, VV17b].
Compartmentalization [LPGB16, AWS+16]. Compartmentalizing [JBMM16]. compartments [CZW+18, CXZ+18, KMB+15, KJ16].
compensate [LTC+18]. compensation [Góm17]. compete [DAT+18, SG17, Sho16y]. competence [WWW+18, YVMS18].
Competitive [BDK+18]. complete [Bob17]. completion [SOP+16]. complex [AHA+19, BSK+19, BPW15, CWG15, CGPB17, CGY+19, CTI+19, CRA+19].
CBF+18, Con16, CSC+15, CBH+15, DOA+17, DQB+16, DWH+17b, EEE+16, FTDC17, GFvA+15, GPS+17, GBM+15, HK15, HHS+16, IBFDB18, JRH+16, KHRL17, KCB+16, KMLG+15, KMLG+16, KJTY+19, KSM+18, LPRW17, LRBB15, Lep15g, LHA+15, LKE+15, cLN+16, LDR+19, LTRW15, Mes16, MKA+17, MDC+16, NNH17, NDRJ15, NGS+16, NIS+16, RPMC+16, RGM+16, SCNTC+18, SZF+15, SMC+15, Sho15c, Sho15-31, Sho15-43, SHO+18g, SKZ+18b, SBC+16a, SBC+16b, SMOO17, SCL+19, TBF+18, TE+15, TRM+16, VGB+17, WSP+18, YL+15, YIT+15, ZNR+18.

complex-dependent [NIS+16]. complexes
[ACRM17, DMD19, KPA+16, KNL+17, LPRW17, LTC+16, LR18, LBJ+19, MSL16, RMGM18, RND+17, Sho15-51]. Complexin [SES+19]. complexity [Sho15c, Spe17b]. component [MCM+17, MST+15, TBJ+17]. components [AKTR+18, CGY+19, MSK+18, SPGB+17, WF15]. composed [MYT+16]. composition [HHS+16, IZB+17, KBB+15, KBB+16, SOK+18].

Comprehensive [HKK+19], compression [KS+17]. compromises [XIZ+18].


cone [BFDD19, CG16, IY+18, WRP+16]. confer [YGM+17]. confers [PTK+16]. confinement [GBD+18, HHI16, SMM+16].

confinement-dependent [GBD+18]. conflicts [UDH+16]. deformation [DTW+16, OBS+17, SMK+18]. Conformational [FSB+15, WHL+17].


Constitutive [HKT+17, KW+15]. constrain [Bro16, LWF+15, MSV+19].

constraining [CLBB15]. constrained [XPZ+19]. constriction [CJS+18, Jan18, MMW+19, RHH+18]. construction [Gen17]. consult [Sho17b]. contact [BDK+18, CMMB+15, FKL+18a, FKL+18b, GRS+18, GKG16, GBM+15, Hen19, KB16, KLHC+18, MKD+18, MYN+17, PHA+17, SA19, SZ17a, SKZ+18a, SDP+15a, SDP+15b, VMR+19].

contact-dependent [CMMB+15]. contacts [AEP+17, DLH+19, DPcS+18, DSS+15, GY+18, MS19a, MS19b, MST+15, SBS+18, Sho15-50, SK18b, SJL+19, VRM+19]. contain [CST+17, KdKvdK15]. containing [LB+19, NDRJ15, TCP+15, ZJM+17, LLW+17]. content [HAK+15, SZS+18]. contractile [KTM19, MSK+18, SOP+16, WMB+15].

contractility [AHA+19, KT15a, KT15b, NWP+16, Wu17]. contraction
[CHP+17, FTAB+15, GKC+17, JhZbYmP15, MXV+16, TY16]. contractions [MRMM18]. contribute [GCZ+19, HNF+18, Mar16a, SFG+17, SKN19]. contributes [LXR+15, SMK+18]. Contribution [NEW+17, VPD+16]. contributions [ECAB+16]. Control [AHs+18, DWH+17a, LLW+15, SPK+18, AZS+15, BCH+17, BSP16, CD18, CBAP+17, Can17, CS16b, CYMS+19, CE16, CED+15, DPGS+18, DSSF+15, DZB+18, FG15, GJFR16, GWZ+19, GSKL+18, GN18, GSM+15, GCC+18, HGC+19, HHM15, HCN+15, HCS+18, HB18, IM16, LL17, LOG15, Les15x, LM19, LZ16, LFK+17b, LVG+18, MKA+19, iNLM+19, NGG+16, PXN18, PLD+15, RLM+15, SSM+18, SZE19, SAF+19, SG18a, SG18b, SLAR+16, SK18b, SB19, TWD+17, UBBSM15, WV18a, YHS+15, YLD+16, vdVFM+17]. controlled [ABF+16, ANM+19, MCM+17, RLJ+17, TJMM+18, WF15, vHGD+15]. controlling [CST+16, DCM+17, DLBMA+15, SPMM+17, SHH+16, WBNH18, WWZ+17]. controls [ALLA18, BHS+19, CW17, CIK+17, CCLL17, CLL+16, CSYB+17, CRA+19, CKKG17, CHB+16, Das17, DCB+15, DLT+18, EVR+19, FBBRCA+18, FVF+16, FC19, GDB+17, GPPI+18, GCJ+15, GCH15, GGL+19, HKH16, HAK+15, HQW15, HPW+17, HKT+17, HDA+17, HAR+15, JPF+16, KCb+16, KKP+17, KSM+18, KQM+19, LSPC16, LLC+17, LTS17, LDG+15, MRGW+16, MdpN+17, MDC+16, NHG+18, OTG+18, OSK+15, PLS+15, PSC+15, PAM+16, PST18, PMG+17, PKKB17, QJP+17, RHPH+18, RBM+19, RS+W+15, SD1+19, SHW+17, SVd+15, SSH+15, SEMP15, SCK+19, SiYM+18, SKZ+18a, SDP+15a, SDP+15b, SYK+17, TCD+15, WYHG17, WZR19, WQD+18, WV18b, ZJM+17, ZDSM+18]. convene [Kon17]. convention [Sle16]. converge [HMC+16, RM+16]. conversion [CW+17, LJS+16a, LJS+16b]. converts [RYS+15, WWY+18]. cooperate [BCS+17, DAG+15, SNOMM16, Sho16y]. cooperates [GWZ+19, LLS+18, RAS+19, SES+19]. cooperating [WRV+15]. cooperation [Mar17]. cooperatively [TAQ+19]. coordinate [CAKL16, CWZ+15, PUTM15]. Coordinated [EVR+19, LZC+15, HTK+16, LCD+17]. coordinately [HBWY18, MKS17, ONT+19, PPK+16]. coordinates [BCM+18, GBRH15, LLIW+17, MF18, MKD+18, NYT+16, RMMS+17, SJJ+19, YSM+17]. Coordinating [Jor16c]. coordination [CZP16, MCGM15a, MCGM15b]. cop [Sho16-27]. COPII [Far16, Gli17, GYK+17, RBP+17, SKN19]. COPII-coated [GYK+17]. COPII-dependent [Far16]. cord [CBAP+17]. core [NNH17, PTK16, ZJM+17]. Coronin [BRY+19, HBDW+15]. corrals [LTG+18]. correct [DLM+15, IKRMN16]. Correction [BP19a, CSF+18, DMH+15, DCO+16, DKR+19a, FLN+16, FLG+19, FCB+19, FKL+18a, GKK16a, GHS16a, KT15a, KMK+17a, KM18a, KMLG+16, KBB+16, KOV+16a, KST+17a, LLAC18b, LJS+16a, MS19a, MSW+17, MWSM19, QSZ+17a, RLS18a, SG18a, SBC+16a, TSK+19, VBJ+18a, WCY+16a, gXNG+16, XRH+18a, HBM+19]. Correlative
Cortactin [GM16, SHH+16, HQW15, KBT+15].
Cortical [JDZ+16, AGL+16, CMB+15, CSA19, DOH+17, GM18, KL17, LM15, LSMG18, NLS+18, NDRJ15, OKN+16, Sho15-33, YVIM18].


curbs [HLST19]. curvature [BJO+16, CWG19, DWH+17a, JDG16, LMM16, McM19, SHR17, XIZ+18]. curved [MOM+18]. cut [Rab17]. Cutting [BP19a, BP19b, CGT16].

cytogenetic-scale [SWD+19]. cyto hesin [RSC+19]. cyto hesin-1 [RSC+19]. cytokinesis [ABP+19, DPS+18, DOH+17, DKR+19b, JDZ+16, LSPC16, MSK+18, Pol17, RBC+17, SOP+16, SWC+17, DKR+19a].
Cytokinetic [Sho16f, BDW19, CHP+17, CWL+16, WG16]. cytological [CZW+18]. Cytoskeleton [HKG+18, SES+19]. cytoplasm [ABF+16, PH16]. cytoplasmic [BYMS+19, BBK16, KDR+19, KJC+15, SFG+17, SMA+19].


D [BSL+15, CZW+18, GTMZ+15, MF16b, NKH+19, NPC17, PHKY17, PSL+17, SPJ+15, TYD+15, VZB19, dFEEW+15]. DAAM1 [NIS+16, YHS+15]. Dam1C [NDC+19]. Dam1C/DASH [NDC+19].

damage [BSP+17, CR18, Gek17, OR17, OLL+17, PKN+15, RZS+15, RM16, SG17, WZC+15, WBNH18, XPZ+19, XTS+15]. damage-induced [Gek17]. Damaged [Sho151, BJ+18, DBS18, PSCS16]. DAX [MBS+17, Inf18b].


Deciphering [FWH+16, O’D16b]. decision [AS17, BOL17, Sho17b]. decisions [HH18]. Decoding [Spe17b]. decondensation [KPGG+19].

deconstructing [PVP18]. decreased [WGHE+18]. decreases [MNL+16].

Decrypting [Sho16g, Cas17b]. deep [Fuc15, GTW+15, SK16a]. Defective [BLO+16, CS16b, ZWB+19]. Defects [AEP+17, Blo19, CNRR+17, MNL+16, OSW+17, RSG+15]. defense [Sed15j].

deficiency [MHG+19, VGB+17]. deficient [BRY+19, CRC+15, WCY+16a, WCY+16b, YKO+16]. deficits [VXF+15, ZYL+16]. define [Sed16c]. defines [BBFD19, MOJ16, RFO+16, WFS15]. degeneration [BBW16, Qi17, WFOA15, WTSA17]. Degradation [BMM+19, gXNG+15, YDM+18, BHS+16, DCM+17, KDM+18, KJTY19, LHI+19, LTB+17, MTTG18, MOS+18, MRK+18, Nic19, PA19, POE+16, PMP+17, SPGB+17, STR+18, SIO+16, TGK+19, UOT+16, WWZ+17, WLJ16, YHG+17, gXNG+16].


demonstrates [SLD\(^{+}\)]]. Demystifying [Sed\(^{+}\)]]. dendrite [KNY\(^{+}\), NC\(^{+}\)]. dendrite-specific [KNY\(^{+}\)]. dendrites [Bro\(^{+}\), GS\(^{+}\)]. Dendritic [Nie\(^{+}\), PM\(^{+}\), BSL\(^{+}\), BJL\(^{+}\), CPC\(^{+}\), CLBB\(^{+}\), FTS\(^{+}\), GSS\(^{+}\), ISL\(^{+}\), LMR\(^{+}\), LLL\(^{+}\), LSS\(^{+}\), OPP\(^{+}\), Qi\(^{+}\), Sc\(^{+}\), SSH\(^{+}\), TTU\(^{+}\), VRK\(^{+}\), WQD\(^{+}\), YDM\(^{+}\)]. DENND2B [IBG\(^{+}\)]. Dense [ASM\(^{+}\), Sho\(^{+}\), NNH\(^{+}\), ZJM\(^{+}\)]. dense-core [ZJM\(^{+}\)]. density [JPC\(^{+}\)]. Dent [Sed\(^{+}\)]. dependencies [Sis\(^{+}\)]. dependent

[ASZ\(^{+}\), AZS\(^{+}\), APHH\(^{+}\), AQL\(^{+}\), AWL\(^{+}\), APS\(^{+}\), ACG\(^{+}\), ACRI\(^{+}\), AIS\(^{+}\), OPL\(^{+}\), CKB\(^{+}\), CKX\(^{+}\), CVL\(^{+}\), CMM\(^{+}\), CRS\(^{+}\), CBM\(^{+}\), DQB\(^{+}\), DLM\(^{+}\), DKA\(^{+}\), DLBMA\(^{+}\), DCF\(^{+}\), EEE\(^{+}\), Fur\(^{+}\), FBPN\(^{+}\), FdAV\(^{+}\), GDD\(^{+}\), GTW\(^{+}\), GBD\(^{+}\), GLJ\(^{+}\), GML\(^{+}\), GLC\(^{+}\), GS\(^{+}\), GWF\(^{+}\), HBBG\(^{+}\), HBDW\(^{+}\), HZH\(^{+}\), JJW\(^{+}\), KT\(^{+}\), KSG\(^{+}\), LAM\(^{+}\), LRH\(^{+}\), LOG\(^{+}\), LMR\(^{+}\), LSP\(^{+}\), LKM\(^{+}\), LGH\(^{+}\), LDP\(^{+}\), MTC\(^{+}\), MPH\(^{+}\), MCG\(^{+}\), MLF\(^{+}\), MF\(^{+}\), MCG\(^{+}\), MDC\(^{+}\), NIS\(^{+}\), OMK\(^{+}\), OMK\(^{+}\), OLM\(^{+}\), QYC\(^{+}\), SPD\(^{+}\), SSL\(^{+}\), SSH\(^{+}\), SDHC\(^{+}\), Sor\(^{+}\), TBK\(^{+}\), WFOA\(^{+}\), WZG\(^{+}\), WW\(^{+}\), WF\(^{+}\), YEM\(^{+}\), YTL\(^{+}\), YMD\(^{+}\), YSR\(^{+}\), ZAT\(^{+}\), ZWZ\(^{+}\), ZCL\(^{+}\)].

depends [CMB\(^{+}\), JNS\(^{+}\), RSC\(^{+}\)]. DepHining [Ham\(^{+}\)].

dephosphorylation [CHB\(^{+}\), LHT\(^{+}\), PS\(^{+}\)]. depletion [BR\(^{+}\), CCS\(^{+}\), GLL\(^{+}\), GLL\(^{+}\), HDA\(^{+}\), MLJ\(^{+}\), PS\(^{+}\)]. depolymerase [BRH\(^{+}\)]. depolymerization [ARV\(^{+}\)]. depolymerizing [VGA\(^{+}\)]. deposition [KS\(^{+}\), LSP\(^{+}\)]. deprived [HSK\(^{+}\)]. derived [AKTR\(^{+}\), ASY\(^{+}\), DMC\(^{+}\), Juh\(^{+}\), SCL\(^{+}\)]. Designing [JW\(^{+}\)].

desmin [ARV\(^{+}\)]. Desmoplakin [KDM\(^{+}\), AZS\(^{+}\), Les\(^{+}\)].

desmosomes [Sho\(^{+}\)]. destined [MRK\(^{+}\)]. destruction [NOS\(^{+}\)].

Detaching [Lac\(^{+}\)]. detachment [ACRM\(^{+}\)]. detect [DRMW\(^{+}\)]. detected [XTT\(^{+}\)]. determinants [EBMW\(^{+}\), UKHK\(^{+}\), VRK\(^{+}\), VRM\(^{+}\)].

determination [LSL\(^{+}\)]. determine [YEM\(^{+}\)]. determined [BYU\(^{+}\), OCS\(^{+}\)]. determines [CHL\(^{+}\), CGB\(^{+}\), HPB\(^{+}\), JOG\(^{+}\), MWW\(^{+}\), PhHS\(^{+}\), RGOS\(^{+}\), SRI\(^{+}\), WX\(^{+}\), WCY\(^{+}\), WCY\(^{+}\)].

Determining [PS\(^{+}\)]. Detyrosinated [MSV\(^{+}\)]. detyrosination [YCS\(^{+}\)].

Deubiquitinating [RD\(^{+}\), YLW\(^{+}\)]. Deusen [Pow\(^{+}\)].

developing [CIS\(^{+}\), DGS\(^{+}\), MRO\(^{+}\), MLR\(^{+}\), SCL\(^{+}\)].

development [DSD\(^{+}\), GJF\(^{+}\), GFH\(^{+}\), GGL\(^{+}\), HGC\(^{+}\), HKG\(^{+}\), HNC\(^{+}\), KVZ\(^{+}\), LLS\(^{+}\), LMR\(^{+}\), LTX\(^{+}\), OW\(^{+}\), PA\(^{+}\), PSC\(^{+}\), RGR\(^{+}\), SM\(^{+}\), Sho\(^{+}\), SCK\(^{+}\), SCP\(^{+}\), SLG\(^{+}\), THG\(^{+}\), TS\(^{+}\), ZWW\(^{+}\), dVGO\(^{+}\), vS\(^{+}\)].

development-specific [GFH\(^{+}\)]. Developmental [SJJ\(^{+}\), DD\(^{+}\), DSSF\(^{+}\), ITN\(^{+}\), She\(^{+}\)].

Developmentally [LHY\(^{+}\), CLH\(^{+}\)].

devices [EWL\(^{+}\)].

diabetes [FBPN\(^{+}\)].

diai-dependent [FBPN\(^{+}\)].

diabetic [CIK\(^{+}\), ZPT\(^{+}\)].

diacidic [CGB\(^{+}\)].

Diaphanous [LM\(^{+}\)]. diaphragms [CRPSC\(^{+}\)]. DICER [CR\(^{+}\), BSP\(^{+}\)].
DICER- [CR18]. Dickkopf [DMC+16]. Dickkopf-related [DMC+16].
dictate [JCK+19]. dictated [SFA+19]. dictates [GB18, Sho15n].
differences [HGL+17, LS18]. Different
[IdSCB+16, DTW+16, HCS+18, LDM17, PKC+16, RGM18]. Differential
[CM18, MSS+17, AB18, BLG+15, GAS+18, OKN+16]. differentially
[GLL+18b, HGC+19, KLHC+18, NLBA+15, ZPT+15]. differentiation
[BSK+19, BMP+18, BMS+17, CWL+17, CSG+15, CEM+15, CRK+17,
DSC+18, DAG+15, EPF16, GCH15, GCC+18, MSL16, OBY+15, PAM+16,
RSCR15, Sed15e, Sho16c, SQB+15, UGB+16, WYHG17, YGW+17,
ZGDS+16, dFEvW+15]. diffuses [RZS+15]. Diffusion
[UKHK15, TRM+16, TG15]. digested [Les16c]. Digging [TG17].
dimensions [SB17], dimer [WMH+18]. Dimerization [Sho15n, WBL15].
dimers [MB17a]. dine [TS15b]. DIP [NWD+19]. DIP-2 [NWD+19].
diploid [YYM+18]. Direct
[VM19, CYT+18, FRP+17, JKA+15, KTK+18, KMJ+18, PBL+19, WMB+15].
Directed [HLW+15, SW18, CEM+15, DRL+19, FLN+10, FLN+16,
NiYT+16, YTTH+17, YVM18, GSD+15]. directing [YGW+17]. directional
[EAW+17]. directionality [CLL+16]. directly
[ALY+17, BBSA+16, DOA+17, SKZ+18b, vBMG+15]. directs
[APS+17, CWI+19, HLHF15, KDA+18, SWS+19, SHR17, TLH+19].
disables [PSC+16]. disaggregase [OCS+15]. disassembles [SAK+18].
disassembly [IBFDB18, LDG+15, WW16, WMH+18]. disc [MG17, Pug15].
Discrete [CAKL16, MTC+19]. Discs [DSA15, FLG+18, SPD+17]. Disease
[HV17, CS16b, DLH+19, HHS18, KJH18, NPC17, SS19, TS15a, VV17b].
Disease-associated [HV17]. diseases
[HPE+19, KM17, KM18a, MB17b, VZ17]. Dishevelled [LHY+19].
disjunction [RSG+15]. dismutases [WBNH18]. disordered
[BA18, SZK+19]. dispensable [SSPD15, SSR+17]. dispersal [SDW+19].
dispersion [RSW+15]. displays [TBW+17]. disrupt [YCS+17]. Disrupted
[Sho16h]. disrupting [ZWW+19]. disrupts [DSH+18]. dissection [MP17a].
dissemination [SSE+18]. distal [DER+18]. distance
[MTC+19, MS19a, MS19b]. distance-dependent [MTC+19]. distant
[BLL15]. Distinct [AFX+16, BPSK+16, BSP16, GDV+19, CGPB17, CCKG17,
DDAR+16, HKH16, JCK+19, KGN+15, LPRW17, LTG+18, MSE+17,
MSL16, RGM18, SSL+17, SPWM15, SHVO+18, VLP+15, WWZ+18].
distinction [LDN+15]. distinguishing [PCK+17]. distorting [TSF+15].
distribution [KST+19, SIYM+18, WZR19]. disulfide
[FC15, Mok16, RPM1+16]. dive [SK16a]. Divergent [MSL16]. Diverse
[ZTR+17, ND17, RNP+17]. diversify [Sho17d]. diverts [MG17]. dives
[Sed16a]. divide [Inf18a, Les15-31]. dividing [Gra16]. Division
[CG16, AZ19, CJS+18, CKX+16, FK17, JDZ+16, Les15a, PUTM15, PSC+15,
SXE+19, UMC+15, UMC+17, VY18, YJF+16, ZB18]. divisions [LDM17].
Dkk1 [DMC+16]. Dlk1 [TGQ+17, TAQ+19]. Dlg1 [AHA+19]. Dlx3
[UBBSM15]. DNA [AWL18, ABGG16, Bob17, BG18, BHS18, BSP+17,
BCMM+19, Can19, CR18, Col18, DLM+15, EMRS+18, Gek17, Ger15, GCA+17, GRB19, GCW+16, LVF+15, LS16, LCD+17, Lov18, MTGG18, MN17, OR17, OLL+17, PMRM17, PMHB17, PKN+15, PUY+19, Pri17, RZS+15, RS19, RLS18a, RLS18b, SG19, Sed15q, SG17, TSFP+15, WZC+15, WSP+18, XIZ+18, XPZ+19, XT5+15, YGMR+17, YTGA16, vV17a. DNA2
[HESKK15a, HESKK15b]. DOCK7 [NYW+17]. docking [GDD+15]. does [PKS+19, SD17, TT19]. doesn't [Les15n]. Doing [NA16]. Domain
[LL17, BPH+18, BA18, CE16, DZL+15, GLC+19, GUM+18, ISL+18, KCB+16, LBB+15, LDG+15, NL16, RC15, SOW+17, SLG+18, TCP+15, WYV+19, ZJM+17]. domain-containing [TCP+15]. Domain-specific
[LL17]. domains [CST+17, HKM+15, KGN+15, KNO+19, PPK+16, SOW+19, SG19, SZK+19, SJJ+19, XRH+18a, XRH+18b]. dominant
[BLL15, PMHB17]. double-stranded [BS+17]. doublets [BMF+18]. down [Inf19b, NL16, PMP+17, vV17a]. down-regulation [PMP+17].
downs [Z16]. downstream
[BS16, GFWG15, NWY15, PSC+15, SYK+17]. DPP
[DKMV15, LWF+15]. DPP-mediated [DKMV15]. Draper [WV18a].
Draxin [HB18]. drift [HKT+17]. drive
[GGF+19, JBE+17, MRMM18, MSE+17, NLBA+15, OKN+16, SMF+15, SSL+17, Sho16h, SZK+19, SBC+16a, SBC+16b, TNP+15, XSI16, YTTH+17].
driven
[CPBG19, FFG+18, HOH+16, KL17, MB17b, SNGO16, LHA+15, PGMM+19].
driver [NKW+19]. drives
[BNB+15, CBB15, DRL+19, GUM+18, KS17, KJZ+19, KSGL19, LRH+15, LCP+15, LEM17, Lin15, LE16, PSL+17, QYC+17, TBL+15]. driving
[WH1L17]. drop [Les15c]. droplet
[Boh18, COGP15, DLH+19, EBMW+18, GBM+15, HSB+19, NO19, OKY+16, SWS+19, SAB+18, VTG+16, XLW+18, DLH+19]. droplets
[CWT+19, FW16, GBK+15, GSB+15, NO19, SMA+19, TJMM+18].
Drosophila
[EG19, SDW+19, TCWM18, TNK18, CKJ+15, CLH+18, DSS+15, DKKM15, FAH+17, FLG+18, KVK+17, KDA+18, KPEJ17, LSMZ+18, LZC+15, LPH16, LL5+18, LHF+15, MBG+18b, OSW+17, POTZ15, RGR+18, RMB+18, RAS+19, SOW+17, TSJ+15, VP+16, WLM+15, XSI16]. Drp1
[Gr16, JCF+17, OMK16, iYJF+16]. Drp1-dependent [OMK16].
[HOH+16, LLZ+19]. druggable [NKW+19]. DSCR1 [CG16, WRH+16].
dTBC1D7 [RHJ18]. Dual
[MSK+19, NYW+17, WWT17, ABP+19, WBNH18]. Duchenne [NWP+16].
due [MSK+18]. Duménil [SIl16a]. Dumont [Pow16e]. duplication
[KMC+19, LUC+15, RND+17]. duration [LK17, MAK+16]. during [ABP+19, AGB+19, AIS+18, BPH+15, BMP+18, BMC15, BVR+17, BDZ+15, BCM1+9, CPM+18, CM18, CHL+19, CMTH+15, CO19, CHH+15, CPH+16, CHS+16, DSC+18, DSSF+15, DB15b, DKS15, DKMV15, EJK+16, EMB+15, EPF16, FLM+17, FDR+16, FMS+19, FCL19, FCL16, GMLT18, GKGK16, GGL+19, GCW+16, HM19, HLHF15, IZZ+18, IZBH+17, JPC+17, JAH18, JDB+16, KV17, KCB+16, KTM19, KPEJ17, KFAM17, Lac19, LLS+16, LPC16, Les15-32, LWH+18, LDP+15, MSK+18, MOJ16, MGO+18, MXV+16, MHG+19, MvV+16, MTGG18, MSW+07, MSW+17, MKD+18, MSK+19, MSL16, NV17, NFYY15, NPU+16, OBY+15, OW19, PUT15, PTR+19, PD19, PSC+15, RB18, SvZS+16, SHV+17, SRT+18, SOW+17, SK18a, SCK+19, SKO+15, SBC+16a, SBC+16b, SNB+18, SCP+15, TYD+15, TF16, UGG18, VRK+17, VTG+16, WMH+18, X16, YJF+16, ZZMC+15].

duties [Les15q]. Dyche [Pow15b]. dye [PK+17]. dynactin [HV17, SV16].

Dynamic [CYMS+19, DJY+16, GM18, GP17, IZBH+17, SMB+19, BMP+18, BP19a, BP19b, CSA19, EGY+19, GTW+15, GKC+17, LJ16, LLL+15, NiG+18, QYC+17, S16a, SSE18, XRH+18a, XRH+18b, CJ16]. dynamically [TM+19].


DYRK [UMC+15, UMC+17].

DYRK-family [UMC+15, UMC+17]. dysfunction [AGG+16, KIN17, KN18a, Pow15a, Qi17, TUE+17, YKO+16]. dysplasia [RMB+18]. dysregulation [PC17]. dystroglycan [CPEE+15]. dystrophy [CMK+16, NW+16].

E-cadherin [BMC15, BKG+15, GBD+18, HLHF15, RMS+18, VHB+18].

E-cadherin/ [BKG+15].

E-catenin [BKG+15]. E2 [CBAP+17]. E2F1 [ZCL+15]. E3 [CHL+19, GCW+16, SvZS+16, SSV+18, WXFS17, WXZ+15]. each [ES18, LB18]. Early [AUT16, JJW17, AIS+18, Góm17, GB+15, HCN+15, LPGB16, LJS+16a, LJS+16b, MP17a, MRMM18, O’D18b, SERP16, SERT+18, SKL+18, Mar19].
epitranscriptomics [YVIMS18]. Epo1p [NDRJ15]. EPS8 [GDB+15]. equatorial [ZCH+18].

extrachromosomally [CPB+16, KMJ+18, SCP+17, DN17, SAO+17, SW18]. extrachromosomally [CPB+16, KMJ+18, SCP+17, DN17, SAO+17, SW18]. extract [GSC+16, PLD17], extraction [SDi+19, STi7]. extracts [RBRi9]. Extramitochondrial [RXEB+19]. extravasate [TCWM18], extravasate [TCWM18], extravasate [TCWM18], extravasate [TCWM18], extravasate [TCWM18], extravasate [TCWM18].

[AGB′19]. filament
[ARV′18, CHP′17, FC15, HM19, ISL′18, KKD′16, LEM17, RBC′17, VQ17]. filaments [DPG′18, ITG′18, Pow15b, RHH′18, WDW′17]. Filamin
[KST′19]. fill [Kaw17, Sho15–73]. filled [FLLM17]. filopodia
[HHT′16, JPC′17, MvV′16, SS16, Sho16–28, UBR′17]. Filopodyan
[BYU17, BCH′17, BPW′17, CRs′17, DBG′15, LCP′15, LXR′15, OMKM16, PSCS16, RHH′18, SPGB′17, SPK′18, Sed15w, Sho15q, Sho16p, SZK′19, SK18b, TBK′16, YAH15, YIT15]. fit [Les15k, Sho16a]. fitness
[RLM′15]. fits [BA18]. Fitting [O′D18c]. Five [MG16]. FKB [APBS17]. FKB-6 [APBS17]. flagellum [BMF′18, RGR′18]. FLCN [MF18]. Fld1
[GBM′15]. Fld1/Lbd16 [GBM′15]. flies [AWS′18, DCO′12, DCO′16]. flight [DSS′15]. FLIM [VRM′19]. flipase [RSvW′15]. flipped
[UFT′15]. flips [Les15t]. flow
[CDT′19, CBB15, FG16, LLS′18, SHW′17, Sho15v, Sed15o]. Flower
[CMB′18]. fluid [BLO′16, LTG′18, Nie16]. fluidity [SDP′15a, SDP′15b]. fluorescence [BDW′15]. fluorescent [BCG′19, KSM′17, PABM16]. fluorescently [DSC′18]. flux [FBX′15, KBJ16, RGOS′16]. Flying
[O′D18f]. FMRP [Log17]. FNIP [MF18]. Focal
[GGF′19, FKG′19, JBE′17, JIB′19, KSG′16, KOV′16a, KOV′16b, MCD′19, Sho16s, SHVO′18, TLMG′15]. foci [DMG′19, SHR17]. focus
[ML15b, Sho15u]. focused [Les16b]. focuses [IG15]. folding
[KTM19, KML′15, LFK′17a, LGH′18, TSK′18, TSK′19]. follicle
[PGM′19]. follicles [GI19]. follicular [dVGO′16]. following
[LUC′15, SWD′19]. Folsch [Jor16g]. force
[ACRM17, BBHBFSP18, FTAB′15, HB16, Jor16h, KTM19, KBT′15, KOV′16a, KOV′16b, KST′19, PLD17, RBZ18, WXC′18]. force-dependent [ACRM17]. force-sensitive [RBZ18]. forces
[AZ19, BGJ′16, DPGS′18, DN16, JhZbYmP15, Les15s, MSLK′18, Nel17, PBL′16, Pow15e, SXE′19, SWC′17]. Forkible [NOS′15]. forebrain
[NYW′17]. fork [BG18, Can19, CQB′19, ZDM′15]. forks
[BCMM′19, Les15i, RS19, RLS18a, RLS18b, TBL′15]. form
[BLZ′15, CPCtH′15, CGY′19, DSA15, LPRW17, SPD′17, Sho15–56]. Forman [CJ16]. formation [BSL′15, BM15, BS17b, CRPS1′19, CHC′18, CPB′16, DPS′18, DS16a, DJV′16, DLZ′15, FFATC15, GPAA′18, GCZ′19, GLL′18b, HKG17, HM19, IYP′18, ISL′18, JSB′18, KHA′18, KBB′15, KBB′16, KST′17a, KST′17b, MSS′17, MHI′18, NWP′16, NPU′16, OKY′16, ONT′19, Ott16, PM15, RMMS′17, SOH18, Sho15m, SLM′15, SENL′15, SAO′17, SDP′15a, SDP′15b, SCL′16, SAB′18, TCD′15, VPD′16, WPA′18, WEQ′15, WMB′15, iYF′16]. Formin
[MHY′16, DBG′15, GTW′15, GSKL′18, LM16, PAC′15, WMB′15,


Francesca [Pow15c]. Fred [Jor16c]. Fredberg [Sed15o]. Frederic [Jor16e].
free [JSB+18, Sho18a, TG15, WFS15]. fresh [Les15i].

FRET [OSL+19, VFM+19]. fringes [Pow15b]. Fritz [O’D19g]. front [JGCAC+15].

frontal [Les15r]. frontotropic [WLM+15]. Fuchs [O’D17g].

Fueling [Yel18]. fuels [KML+15]. full [GK+18, Les15r, LDR+19]. full-length [GKG+18, LDR+19]. function [AEP+17, BKH+15, BC19, BMC15, BCM+19, CKJ+15, CQB+19, CCS+19, CWL+16, CSYB+17, CCY+19, DMC+16, DKS+15, DB15a, DCF+17, EJ+16, FLM+17, GFLA+15, GLL+18b, Gen17, GGC+17, GYK+17, HGD+15, KHR17, KTK+18, KSM+18, LJP+15, Les15z, LT19b, LTRW15, MCS+15, RXEB+19, RSwV+15, Sed16a, Sho15-56, Sho16-36, TMK18, WZC+15, WYV+19, WF15].

functional [CKS+15, CN15, NBG+16, NGE+16]. functionally [ML+16].

functions [ATRG19, BKG+15, CD18, DsvNA+15a, DsvNA+15b, FLG+15, FLG+19, ML1+16, MRM18, MGT+19, NNN+15, OSK+15, PLD+15, SPPD15, TAQ+19].

Fundamental [Sch15].

fungal [VAB+18].

Furin [BMC+15]. furious [GB18].

furrow [FLN+10, FLN+16, LW16b, PUTM15]. furrows [WG16]. FUS [MCH+18, YCSJ+17]. fuse [BUPC19]. Fusion [BCH+17, BPL+18, CZZ+15, CRC+15, CQL+15, CLO+19, DOA+17, DSS+15, D16b, DBG+15, FR16, GRU18, GHKW+19, LKM+15a, MPH+15, MJN+18, MSW+07, MSW+17, MKD+18, MS+19, NPU+16, RHPH+18, Sed15m, Sho15-44, SKL+18, SHR17, VML+17, WTC+19, WHL17, WMH+18, YSW+15, vGWC+18].

fusogenic [VK+15]. fusogens [VM+17]. futile [AZ19]. future [SKG17, Tar15]. fuzzy [CBF+18].

Fyn [FBS+15]. FYVE [TCP+15].

G [BSP16, CNC+18, FDs+17, IdSCB+16, LMPG+15, MMW+19, PhHS+16, Sch17a, TLMG+15].

G-protein [TLMG+15].

G0 [B19].

G1 [MAK+16, PKN+15].

G2 [WV18b].

G3BP [KPA+16].

G3BP1 [TT19, ACG+15, PKS+19, SENL+15].

G3BP1-S149 [TT19, PKS+19].

GABAergic [CBAP+17].

GABARAPL1 [SSRG18].

Gabriel [O’D19c].

Gag [HBS+15, Sho15].

Gage [Silt16].

gain [WWW+18].

galectin [KSGL19].

galectin-9 [KSGL19].

game [SG17].

gamete [VML+17].

ganglion [IKR+16, SZL+16].

gap [KDM+18, Sho16f, SOP+16, CM18, TAQ+19].

gastric [ZLG+15].

gastrulation [Pow16c].

Gatekeepers [PW19].

Gatekeeping [Col18].

gating [RPMC+16].

GATOR1 [MF18].

GATOR1-dependent [MF18].

Gauging [Sho15].

GCN2 [KV+17].

GCS1 [VML+17].

GEF [ANM+19, DKS+15, FDR+16, Nie16].

GEF-H1 [FDR+16].

gender

Genetic [MP17a, SSdLA+15]. genetically [MCH+18]. genetics [Pow16d]. Genome [DSC+18, MHI+18, SIBM17, CNA+17, CZW+18, HSN+16, LUC+15, NHA+19, NPC17, UOT+16, VZB19]. Genome-edited [DSC+18]. Genome-wide [MHI+18, SIBM17]. genomes [CSF+17, CSF+18, SZ17b].


gets [Les15p, MB17a, RMTR17, Sho15l, Sho16q].


GLUT4 [BBC+16]. glutamate [FV17]. Glutamylation [MH15].


Going [MS19b, FC16, Les15r, MS19a]. Goley [O’D17b].

Golgi [CPBG19, CGPB17, CBM+16, GNM16, KYN+18, KOK+19, LLL+15, LCTP17, MSCS19, MHA+19, SA19, Sed15p, VRM+19, WDM+15, YWdH+17].

golgin [LLL+15]. González [Sed15b]. good [Les15a].

GOP [SLH17, YHG+17]. GOP-1 [SLH17, YHG+17]. governing [CRK+17]. governs [AUTM16, CXX+16, MBS+18, NVP17].

Gp135 [SHO+15-74]. gp210 [GCH15]. gp210/Nup210 [GCH15]. GPCR

[AMS+17, GAS+15, LL17, MMW+19, TCWM18, VBJ+18a, VBJ+18b]. GPCR-independent [MWV+19]. GPCR-induced [GAS+15]. GPCRs [Les16, YNN18]. GPER1 [BLZ+15].

GPI [LFT+16, LGH+18, Sed15d, SLAR+16]. GPI-anchor [LGH+18].


granule [ACG+15, CMB+18, GDD+15, KPA+16, PIA16, PKS+19, Sho16-31,

hereditary [AEP+17, BLO+16, XTT+18]. herpes [Nie19, TGK+19]. herpesvirus [CSF+17, CSF+18]. herpesviruses [SZI+17].

Heterochromatic [DMG+19, SWD+19]. heterochromatin [MS19a, MS19b, NKH+19]. heterodimer [MF18, dFEV+W+15].


HGF-induced [RSC+19]. HHIP1 [HGD+15]. hi [O’D16a]. Hierarchical [BMW+18, KSL+17]. High [BCG+19, PCF+19, BZG+17, CDF+18, DWH+17a, FGR+18, GPD+19, Pow16d, XIZ+18]. high-avidity [GPD+19]. high-probability [BZG+17]. High-resolution [PCF+19].


HSP90/R2TP [MCM+17]. HSV [JNW+15]. HSV-1 [JNW+15]. Hu [Pow15]. hub [GYS+18, KSL19, VZFG+18]. Human
[NAFM+17, BSK+19, CS16b, CEM+15, DSC+18, GKG+18, HV17, IZZ+18, JNW15, KY15, KWB+15, LBG+17, LBV+17, MB15, MJSB16, MWF+15, NTT+15, NWH+16, NPC17, PTK16, PBS+16, QYY+16, RBC+17, RDN+19, SRT+18, TST+17, THG19, TBL+15, UDH+16, WHP+18, WPA+18, YSR+18, ZDM+15, ZGDS+16, ZCH+18]. Humanin [GTD+18]. humans [SZ17a].


Increased [UGHB+16, CMTH+15, JPC+17]. increases [DOH+17, LRBB15, LMC+18]. increasing [WCY+16a, WCY+16b].

Incredibly [SZ17a]. Independent [IKRMN16, DBC+15, FVF+16, GKGK16, Gra16, JKD+19, KML+15, KJON+17, MMW+19, MBC+19,
Integrin-mediated [SGF16], Integrins [JCK15, KL19, LCZ16, LM19, PSG15, RLM15, SCK17, UOT16].

intensities [BDAW15], intentional [Gar15b], interact [INLM19, PHA17], interacting [AHS15, LRBB15, HBS15].

Interaction [FKO18, CSG15, CKS15, DsvNA15a, DsvNA15b, DLZ15, EGI9, KBT15, LHY19, LSS15, MKA17, PMRM17, PKKB17, SLW18, SCG17, Van19]. interactions [ACG15, BG18, DNM16, FRP17, HQW15, KSM17, MBR19, QZY19, RLS18a, RLS18b, Sho18a, SEMP15, XLW18]. interactome [AR15, JSB18], interacts [HGD15, MCH18], intercellular [AGL15, HVH19, KTM19, SDW19, SZR15], intercellularly [FSF15].

interchangeable [[FFÁTC15], Interchromosomal [MBR19, FMS19, RHC516], interdependence [VZB19], interface [GNM16, NDC19, UFT15], interfaces [PHA17], interferes [MCSC15], interferon [CHZ17, HGG17, WBL15], Interleukin [TCP15].

Interleukin-4 [TCP17], interlocks [MGSO18]. Intermediate [DPG18, FC15, GDV19, LH19, LEM17], intermediates [KMBO15].

Intembrane [MRWM18], internal [BHB18], internalization [CMM15], internalized [JPD16], interneuron [NYW17], internodal [EVR19], interorganelle [HZB19, Hen19], Interphase [LIP15, PSP15], interplay [NP15], Interrogating [BOL17], intestinal [CNC18, KPEI17, RMB18, RRM17, Sho17g, TSJ15, YLND16].

intestine [LRH15], intra [AFXS16], Intracellular [SZL16, CWL17, CSM17, GYK17, MSE17, SPH19, SLYM18, SDW19, UGB16, WGHE18], intracentromere [LVF15].

 intraflagellar [BMF18, YSM17], intrakinetochore [MHA16].

Intranuclear [BPW15], intravacuolar [RNPH17], Intrinsic [MHG19, MGT19], Introducing [NA17], intrinsic [POTZ15], Intronless [WWW15].

inturned [YHS15], invadopodia [ASM15, GLL15b, RHC16, Sho15m], invadosomes [CLO19, POE16].

invagination [MFVS15, SK16a, UBB15]. Invasion [ACG17, CC19, DC17, FBPN18, GML16, HBB17, LR18, MBS18, MCCL15, MBS17, PAC15, RHC16, TB16, TYD15]. involved [BPH15, IsDSC16, SL17], involvement [CBF18], involves [SMOO17]. ion [UFT15, Sed15], IPS [MT19]. IQGAP1 [BK15], IRE1 [BMM19, TCP18, TSK18, TSK19], IRGM [KJF18], iron [DNMB16]. irreversibility [DSSF15], ischemia [LVW17], ischemic [ZZW17], islets [ECL16].

ISM1 [OWW19], isn’t [Sed15q], isoform [CRPSC19, CBF18, KNPC16, OKY16]. isoform-specific [CBF18], isoforms [MSE17, PMR17, TJM18]. isotropic [KST19]. isotype [Kaw17, PTK16], isotype-specific [PTK16], itch [GD16], itself [CSF17, CSF18]. IV [CPB16, JCK19]. Ivaska [Pow15e].

Kinesin-binding

Kinesins [BFDB18, BDLB15, CKKG17]. kinetics [CHB+16].

Kinetochore

KMLG+15, WF15, ASZ+18, BHS+19, CRZ+16, DW17, DRL+19, DRMW17, DMB+18, DSL+17, DUL+19, EG19, GCL+15, GHS16a, GHS16b, HAPC+19, LHB+18, MHSD+15, NDC+19, SPGB+17, SSDLA+15, SD17, Sho15k, Sho16o, VGY+17, YAHH15, ZYA+17, ZGZ+15, KMLG+16]. kineticochore-directed [KMLG+19]. Kinetochores [KMD17b, NIS]. Lattice [LHB+18]. Latrophilin [LHB+18].

Latrophilin-2 [LHB+18].

Kiss [MBR19]. KKT4 [LHB+18]. KLC1 [JERL+15]. KLP [CSC+15].


Kon-tiki [LPHH16]. Kornblihtt [Cas17a]. Köt [Jor16a].

Kras [FdSR+17, WXZ+15]. Krs [Rab17]. Kuduk [DWH+17b].

Kuzbanian [DCO+12, DCO+16].


Latrophilin [AMS+19]. latrophilin-2 [AMS+17].

Latrophilin-2 [AMS+17].

Layers [Jor16g]. Laylin [O’D19g]. LC3 [HM19, Mar16a, NPU+16].


Legionella [AKTR18]. length [AKD+17, FHX+15, GDB+17, GJFR16, GKG+18, LDM15, LRS+17, LDR+19, SCK+19]. Lennon [Sil16a].

leupaxin [KBT+19]. level [LLW+15, MN17, SBM17]. levels
[Far16, GJC+15, GSM+15, LJS+16a, LM16, LJS+16b, MMB+15, NHG+18, NCV+16, SVD+15, Sho15-68, Sho17k, SKZ+18a]. Leydig [GLL+18a]. LFA
[CLW+17, CBB15]. LFA-1 [CLW+17, CBB15]. Lgl [DZL+15]. Lgr5
[CNC+18]. Liaisons [CG17]. Liang [O’D18a]. licenses [Ott16]. licensing
[Blo19, Col18]. life [O’D17g, Sch15, Zha19, Jor16]. lifting [Sho16d]. ligand
[GK+18, HGD+15, WWZ+18]. ligand-induced [GK+18]. ligands
[LWF+15]. ligase [BHS+16, CHL+19, HESKK15a, HESKK15b, HSN+16, LKE15, PNE+19, SZE19, SvZS+16, SVD+15, SMA+19, WXFS17, WXZ+15]. ligases [GCW+16, SSV+18]. Light [WS18, FGR+18, FML+17, GDV19, HYC16, LDMW+15, MFVS18, OSL+19, RMO17]. light-sheet
[FGR+18, OSL+19]. lighting [O’D19c]. like [CNA+17, DMC+17, DVS+17, FFG+18, GFWG15, GUM+18, HKM+15, HR16, ISL+18, JERL+15, KJZ+19, KGN+15, LgYL+18, OG16, SS19, VZ17, YSW+18]. Lillian [O’D19g]. LIM
[BPH+18]. limit [CSC+15, MCD+19]. Limited [SLAR+16]. limiting
[LSMG18, LMDM+16, dVGO+16]. limits [Sho15-61, Sho16a]. LIMK1
[LZD+16]. LIMK1/cofilin [LZD+16]. LIMK1/cofilin-mediated
[SJ16, CWG15, CGY+19, DWH+17b, LTC+16, SR17b, WSP+18]. LINC-ing [SJ16]. Lindquist [Bev17]. line [SHW+17, OO18]. lineage
[CSS+18, MLR+16]. lineage-specific [CSS+18]. lineages [ZGDS+16]. Ling
[O’D17d]. Ling-Ling [O’D17d]. link [GCVAG+18, NS18, Sho15-60]. linkages
[PUY+19, SRZ+15]. linked [EBMW+18, CGBD+17]. linker
[AHS+18, AIS+18, KQM+19, SERP16]. linking [ZAT+17, BBW16]. links
[CSO+19, DN17, HPE+19, OLT+19, PTMP+15, Qi17, SLW+18, VMP16, WZC+15, YHS+15]. Lipid [FW16, MCGC+15, SWS+19, VTG+16, AFO+16, Boh18, CWI+19, COGP15, DLH+19, DJV+16, EBMW+18, GBK+17, GY18, GCL+19, GSB+15, HSB+19, HCC+17, KTK+18, KBB+15, KBB+16, KLIC+18, Mes16, NO19, OPP+18, OKY+16, PKC+16, SII18, Sho15-60, SMA+19, SAB+18, TJJM+18, VTK+15, XLM+18, GBM+15, NO19]. lipid-dependent [GLC+19]. lipids [Kti19, TG15, VYB+19].

[SWS+19]. Lipoprotein [Pfe16]. lipotoxicity [LCTP17, TCZ+16]. liquid
Listeria [SPH+19, ZB19]. LITE [FGR+18]. little [BH15]. Live
[BPS+15, EGY+19, HBS+15, CDF+18, MSA+17, PBS+16, RZS+15, XR1+18a, XRH+18b, SK16a]. Live-cell [HBS+15]. lived
[DLR+19, TALR+19]. liver [ZSH17]. lives [Pow15c]. living [DSL+17, DMH+15, FWH+16, FJ17, LT18, MTH+16, Pow16e, SKG+16, VM19].
[WG16, KJ16, WWT18]. Localization
[ESS+17, AOL+18, GDB+15, HAPC+19, HGD+15, HBDW+15, KYN+18,

metabolically [TMJM18], metabolism [BS18, CSO19, DN17, L216, MSV16, O‘D18c, RGOS16, SPMM17, Yel18].

Metalloprotease [SAF19], Metalloprotease [EWL16, GML16].

metaphase [CO19, DMB18, FBX15, LWZ19, OM19, YK016], metastasis [DCM17, HOH16, Les15-28, Lin15, QCC19].

metazoans [RGMM18], Methionine [LHT19].

methods [JW19].

methylation [CAA17], mevalonate [HOH16].

Mex67p [SLD15+], Mex67p-deficient [CRC15].

MgalA [TMG15].

MHCII [OPP18].

Mia2 [SNOMB16].

Microglia [Gar15].

Microglia [CBAP17, HHS18, Pow15c, WYHG17].

Microglia [HSZ18].

Microtubule [DLBMA15].

Microtubule-dependent [CRC15].

Microtubule-organizing [Sed16a].

Microtubule-organizing [Sed16a].

Microtubule-severing [MRM18].

Microtubules [KTM19, MOM18, SRT18, BNS17, BCM18, DRL19, FKG19, HGL17, IG15, KEV17, LGV18, MSV19, QYC17, RFO16, SMK18, SKZ18b, VGY17, Ver18, VGA15, WWTF17].

Microvilli [LSN18].

Midbody [Ott16, BRACA16].

Midzone [IBFDB18, LDG15, PCF19].
mobility [CMMB+15, CLBB15, RZS+15]. model
[ASPY+16, BFS+19, BYUJ17, CKS+15, DV16, FGR+18, NWP+16, OSW+17, Pug15, War15, WLM+15]. modeling [BPS+15, THG19]. models
[HTLG18, MNL+16], moderation [NW19], modes [IKRMN16, JCK+19], modifications [AZS+15, L116, Pri17, STF18]. Modifiers [HCN+15].
modifying [PLH18, Sed16c], modular [CED+15], modularity [FLS+16].
modulate [DBC+15, vBMG+15], modulated
[DNMB16, DUL+19, LK17, VLP+15]. modulates [BMS+17, CCS+19, DWH+17b, EMB+15, EPP+16, FTAB+15, GDB+15, LFT+16, LHY+19, LKM+15b, MCS+15, MH15, SSV+18, SAT+17, YYY+15, YAH15, ZYA+17].
Modulating [VPV18, XS18]. Modulation [HHS+16, vLvdKR18, PAM+16].
modulator [DSvNA+15a, DSvNA+15b, RGOS+16], modulatory
[VB1+18a, VBJ+18b]. module [GGA+17]. Molecular
[BYUJ17, CLH+18, DSSF+15, GPS+17, LPHH16, VGY+17, VRM+19, BPS+15, CSS+18, CST+16, CO19, FCLo19, GFH+16, Gar15a, HGF+18, HB18, JDG16, MRM18, MKA+17, SPWM15, S117, SGB+17, Tra18].
molecule [CPEE+15, MSvO17, Ndg+18, SPJ+15]. molecules
[EW18], monocyte [NLH+19], monomer [BKG+15]. monomers
[LMR+17, Sch17a], monoubiquitinated [EMRS+18]. Moreno [O'D18g].
morphine [CYH+16, Sholo16], morphine-induced [CYH+16].
morphogenesis [AUTM16, BPH+18, CRA+19, DKMV15, ECAB+16, GGF+19, HBWY18, KS19, KQM+19, MOJ16, MXV+16, MG17, OWV+19, PMRMS17, Pow15], Pug15, RBZ18, SK18a, TST+17, WSDL17].
morphology [BFPD19, HCS+18, SBM+19, TBJ+17, YEM+19, YWdH+17]. Mosaic
[Bea16, KHS+16, SK19], mosaicism [TALR+19]. mossy [LZH+18].
most [Sed16d, SRT+18]. Mother [Ver16, BPSK+16]. Mothers [Sed15b].
motif [CGBD+17, JGCAC+15]. motile [GBS+15]. motility [BFPD19, EMB+15, FLLM17, GHKW+19, HBDW+15, HV17, LM15, LE16, MCD+19, NIS+16, SAT+17, THA+16, TB16, TG17, UFT+15, YBZ+18, ZDS+18].
motion [AWS+16, JPD+16, NTT+15, TKM16]. motoneurons
[MNLB16, MSS+17]. Motor [FFG+18, MGW18, YVM18, zLSSS+18].
motors [Ava18, JER+15, KDR+19, LEM17, SMF+15, S117, YBZ+18].
mounts [Sho15-65]. Mouse [PBG+15, CSG+15, PGB18, SCh+18, SFZ+17].
move [Jor16i, Kel16, LE16, SERP16]. movement
[ABPS17, HLST19, MGSO+18, MH+16, PCF+19, SHW+17, SJ16].
Moving [Gar15a, SR7b]. MOZART1 [cLNF+16]. MPB [SDHC17].
MP51 [HAPC+19, HBM+19, IWM+16, MF16a], Mps3 [KJT19]. MRCK
[GPS+18]. MreB [TLMG+15]. MRN [ABGG16]. mRNA [ALY+17, ADBST+15, AGC+15, BMM+19, BYMS+19, BMS+17, EMRS+18, HCN+15, NC+16, PBL+19, PMP+17, SBR+15, SPJ+15, SLD+15, TTC+16, XJS18].
mRNAs [KNL+17, PH16, SPMM+17, Sed15n, Sho15-38, WWF18].
mRNP [KP18]. mRNPs [BPW15]. MRTF [FBBCB18, HHBG17].
Msd1 [YIT15]. Msi [YLND+16]. Mps [CKX+16]. Msp-dependent


[ZDSM+18, CBF+18, FG15, TZC+15]. Numb/p53 [TZC+15]. number
[CSC+15, FAH+17]. numbers [ZSH17]. Numb'ing [KR18, Nup132
[YAH15]. Nup2 [SMO17]. Nup210 [GCH15]. Nup60 [Les16h]. Nup82
[GFvA+15, Sho15-43]. NuRD [GCA+17]. nutrient [TM18],
nutrient-sensing [TM18]. nutrients [LK17]. NXF1 [BYMS+19, BPW15].

O [Inf18c, BH15, KML+15, LLW+15]. Obelus [VLZ15]. obligatory
[KMBO+15]. observation [HBS+15, VMI19]. Occluding [CPP+18]. occurs
[HHS+16, KD17b, MSW+07, MSW+17, iYJF+16]. octameric [NAFM+17].
off [Sed16a, SFA+19]. off-rate [SFA+19]. offers [FGR+18]. oh [PCM16]. old
[SD17, SK19]. olfactory [KHS+16]. oligodendrocyte
d[IFEvW+15, vBMG+15]. oligodendrocytes [EVR+19]. oligonomic
[RZG+15]. Oligomerization [GPPJ+18, CBM+16, JCF+17]. oligomers
[HBS+15]. oligosaccharyltransferase [SCG17]. Om45 [WLJ18]. OMA1
[KMRD+16]. omics [QPZ+17]. on-site [PAM+16]. Oncogene [MTC17].
Oncogene-inducible [MTC17]. Oncogenic
[RRM+17, GI9, PGMM+19, YGMR+17]. One [BA18, MSvO17, Pow16b].
only [ISL+18, NA16, MA17]. onset
[HBS+15, KBKW19, KMLG+15, KMLG+16]. oocyte
[BCM+18, CSC+15, KBKW19, LTC+18, LWZ+19, LLS+18, PMRM17, Ver18].
oocytes [BBS+17, BTV16, BPSK+16, BCS+17, CO19, DRMW17, GCL+15,
LJ17a, RO18]. oogenesis [O’D17e]. Ooplasmic [LLS+18]. OPA1
[KMRD+16, Les16c]. Open [BDAW15, Ewe18, UBR+17, VAB+18].
Open-source [Ewe18, UBR+17]. Opening [THG19]. operated
[CCQ+18, RYS+15, SBP+16, WWT18]. Op1 [HGF+18]. Opportunities
[HTLG18]. Opposing [KOIT+16, Pow15c]. opposite [KDR+19].
opposite-polarity [KDR+19]. optic [CED+15, Les15]. Optical
[ZZMC+15]. optimal [LT19b]. optogenetic [AHS+18, BOL17].
optogenetics [GGA+17]. Orai1 [SBP+16]. orange [PCK+17]. orchestrate
[BLG+15]. orchestrate [AIK+16, TSFP+15, XSI18]. order
[BD19w, WZR19]. ordered
[DB15b, KP18, MBG+18a, SDP+15a, SDP+15b]. organ [OWW+19, SK18a].
organelle [BDK+18, SJ+19, SPWM15, YWW17]. organelle-exclusion
[SPWM15]. organelle-specific [BDK+18]. organelles [MPMP16].
organism [FWH+16]. organismal [SBM17]. organisms [FGR+18, War15].
Organization
[HR16, AGL+15, BDZ+15, CZW+18, CBB15, Con16, DZB+18, FC15, GFH+16,
KQM+19, LJP+15, NLBA+15, SSM+18, UGG18, WMK+16, YWdH+17].
organize [AFO+16, MKS17]. organizes
[BZG+17, CKJ+15, LFK+17b, MXV+16, RDO+15]. organizing
[TST+17, Sed16a]. Organoids [SB17, HTLG18, MTC17, RRM+17]. organs
[KDA+18]. orientation [EMB+15, FC19, JKA+15, LDM17, LSGMG18].
origami [O’D16b]. origin [Blo19, SCP+17, TGJ+17, Yud19]. origins
[Hyr15, KD17a]. orofaciiodigital [RDO+15]. ORP5 [SKZ+18a]. ORP5/8

P


Passing [O'D18d]. passive [TRM+16]. patch [DPS+18, Sho15-36].
p athogenesis [JERL+15]. pathogenic [GLS+15b, GLS+15a]. pathology [GWF17]. pathway [BHS+16, HOH+16, IGK+16, KVK+17, KP18, KG15, KBT+19, MBF17, MNLB16, MSW+07, MSW+17, MWSM18, MWSM19, MBC+19, MNM+15, NOS+15, OG16, PYO+18, RO18, SYK+17, SB19, TJF18, TSK+19, TSK+19, vLvdKR18]. pathway-driven [HOH+16].
phosphatase-independent [JKD+19, MBC+19]. phosphatases [Nil+19].
phosphate [HHM15, Les+15a, LJ+16a, LJ+16b, VMR+19]. phosphatidic
[HGF+18, VK+15]. phosphatidylinositol
[GCJ+15, HHM15, JJB+19, LJ+16a, LJ+16b]. Phosphatidyserine
[MWT+16]. phospho [WF+15]. phospho-dependent [WF+15].
phosphofructokinase [WD+17, ZSH+17]. phosphofructokinase-1
[WD+17]. Phosphoglycerate [QSZ+17b, QSZ+17a]. Phosphoinositide
[LMR+17, SWC+17, CW+17, DCF+17, WIS+17].
phosphoinositide-activated [WIS+17]. Phosphoinositide-dependent
[LMR+17, DCF+17]. Phosphoinositide-mediated [SWC+17].
phosphoinositides [DWH+17a, DJ+16, O’D18e]. phospholipase
phosphorylate [TAQ+19]. Phosphorylated
[OK+15, BSP+17, HBDW+15]. phosphorylates
[DM+18, TNP+15, UMC+15, UMC+17]. phosphorylating
[HS+18, LRBB+15]. Phosphorylation
[LL+19, PK+19, SCNT+18, TCP+18, TT+19, CDF+18, FBX+15, IKK+18,
JPF+16, LSPC+16, MFVS+18, MNL+16, MBG+18a, MWW+16, OSK+15,
SKW+19, Sho+16b, TGQ+17, TTV+17, WFOA+15, WTS+17, WYW+18,
WKM+15, WV+18b, XMJ+19, Yel+18, YTGA+16]. photoactivated [TCZ+16].
photobleaching [FR+18]. Photoreceptor
[Pug+15, SPD+17, DER+18, HKG+18, MG+17, SPE+17a].
photoreceptor-specific [DER+18]. photoreceptors [DAS+15].
photosynthesis [Pow+16d]. Physical [CK+15, ST+17]. physiological
[DT+16, PPB+15]. PI [CCLL+17, Di+17, GPD+19, HQ+15, KSGL+19,
RSC+19, Sho+15–45, SKZ+18a, WX+18, Yud+19]. PI3 [JJW+17, LLL+18].
PI3-kinase [LLL+18]. PI3K [GPD+19, KG+15, MDC+16].
PI3K-dependent [MDC+16]. PI4P [SE+19, SKZ+18a]. PICK1 [FRP+17].
Piddini [Pow+16b]. PIDDosome [APS+17]. PIDDosome-dependent
[APS+17]. pieces [Sho+18b]. Piecing [Sed+15w]. PIK3CA [RRM+17].
PINK1 [MLMF+16, NPU+16, PSC+16, VGB+17, KPE+17, LSMZ+18, NS+18].
PINK1/Parkin [MLMF+16, NPU+16, NS+18]. PINK1/Parkin-dependent
[MLMF+16]. Pioneer [Tra+18, KS+19, Pow+15d]. pipeline [GSC+16, UBR+17].
PIPs [Yud+19]. piston [PHK+17]. pit [MFVS+18]. PIW1 [ABF+16]. PIX
Pkc1-mediated [vDMR+19]. Pkl1 [YIT+15]. places [MRWM+18].
Plakophilin [DKA+16]. Plakophilin-2 [DKA+16]. planar
[KZ+18, iNLM+19]. plant [BLPV+17, NLS+18, YTT+17]. plants
[DMC+17]. plaque [GWF+17]. Plasma
[RS+16, AKTR+18, BJO+16, DQB+16, Di+17, DZ+15, EEE+16, GDD+15,
GC+15, GPD+19, HHBG+17, MPH+15, MCGC+15, MHA+19, PD+19,
PK+16, RB+16, RS+15, SKZ+18a, TG+15, Yud+19]. plastic [Sil+16b].
plasticity [BSL+15, BLZ+15, DSL+17, FAH+17, LMR+17, LZD+16,
NGG+16, PNE+19, SVD+15, WBL+15]. plasticity-induced [BSL+15].
potential [SSL+17, SM18, ZGDS+16], potential-dependent [SSL+17],
power [Les15-29, vV17a], powerhouse [OI18a]. PP1 [LSMG18],
pre-chylomicrons/VLDLs [SNOBM16]. pre-mRNA [TTC+16].
premRNPs [BPW15]. premRNPs/mRNA [BPW15]. prepares [KHA+18]. Prepatterning [NVP17]. Preprotein [CST+17, BHB+18].
preribosome [BPH+15]. Presenilin [EPF16]. Presenilin-2 [EPF16].
presentation [YZY+15]. presenting [ST17]. presents [DJV+16].
preventing [KMRD+16, ZLZD16]. prevents [DPS+18, PMRM17, RSG+15, VXF+15].
Primary [VAKB+18, ALLA18, ASPY+16, BRACA+16, GDB+17, JNW15, LTG+18, MFP17, OTG+18, Ot+16, PhHS+16, PSL+17, PM15]. prisms [Ger18, ZGDS+16]. priming [FSB+15, KCB+16, KMK+17a, KMK+17b].
primordial [CAK16]. principles [LDG18]. Prion [Hkm15, OCS15, GUM+18, HKG17, VZ17]. Prion-like [Hkm15, GUM+18, VZ17]. prions [Sed15c]. PRMT [AZS+15].
PRMT-1-dependent [AZS+15]. pro [CKM+16, SG17]. Pro- [SG17]. pro-regenerative [CKM+16]. Proactive [GML16]. probability [BZG+17].
Problems [MB15]. process [DB15b, GCZ+19]. processes [BSP+17].
processing [AS17, CYT+18, GHS16a, GHS16b, KMRD+16, LFT+16, LGH+18, RS19, Sho15-49, TTC+16, TBL+15].
progenitor [CSS+18, CLL+16, FGI15, LJI17b, SCP+17, YVIMS18, dlFEvW+15].
progenitors [TGJ+17, WYHG17, WRGB+15]. program [DAG+15, GCVVAGS+18, PS16, Sho16w]. programmed [CLH+18].
programs [HCS+18]. Progranulin [CB16, Sho15-47, NBC+16, ZSDO+15].
progression
[BS18, BBHBFS18, DKR+19a, DKR+19b, HGC+19, JPC+17, Les15c, LLW+15, NKW+19, OSI+15, SLW+18, SENL+15, SNB+18, WWTF17].
progressively [ACRM17]. proinsulin [FWL+17, TSK+18, TSK+19].
projection [ADBST+15]. proliferating [IGK+16]. proliferation
[BP19c, CAKL16, CM18, CRA+19, Col18, FC19, HBWY18, KPEJ17, LJ17b, MLR+16, PLG+15, SBS+18, TSJ+15]. Prolonged
[GMT18, LDU+16, LH15]. prometaphase [SRT+18]. promote
[BVR+17, Bro16, BJL+18, CZZ+15, CQB+19, CLO+19, DSS+15, DSB6, DCO+12, DCO+16, EAW+17, GLC+19, GCA+17, HMC+16, JDZ+16, KTM19, LTC+16, LHT+19, LZC+15, LLW+17, LWH+18, MAJ+17, MNLB16, MCL+15, NLS+18, OSI+15, OPF+19, Pri17, Sho17, SWPS+19, SKL+18, SNC+17, SCL+16, VYB+19, WTS+19, WSC+19, ZNR+18, ZGZ+15].
promoted [HLEM+18]. promoter [GCL+15, WWY+18]. promotes
[APHH+19, ALY+17, ACG+15, ARV+18, BKH+15, BSL+15, BAGM17, BS17b, CIS+17, CLBB15, DFB+18, DBC+15, DLH+19, DKA+16, FdSR+17, FKL+18a, FKL+18b, GDD+15, GBK+17, GHKW+19, GWF17, GHS16a, GHS16b, HSZ+18, IZZ+18, IBG+15, IYP+18, ISL+18, KBKW19, KMG+15, KMLG+16, KMC+19, LAMACE+17, LFK+17a, LNS+19, LLL+18, LTB+17, MMW+19, MF17, MBS+17, MBG+18b, NHCB15, OBY+15, PNE+19, PAC+15, PBL+16, PSP+15, QSZ+17a, QSZ+17b, QZY+19, RGM+16, RMB+18, RKK+18, SCNT+18, SXT+16, SSSG+18, SQC+16, Sho15-44, SOW+17, SHH+16, SCP+15, SNM+16, THA+16, TSJ+15, UD+16, VKT+15, VGB+17, WMB+15, WMH+18, Woo18, gXNG+15, gXNG+16, XJQ+17, XLW+18, YTL15, YKO+16, YHG+17, ZCL+15]. promoting
[ECAB+16, GHF+16, HNF+18, PTK16, ST17, WRV15]. prophase
[PTT+19]. Proasposin [ZSDO+15]. prostaglandin [CBAP+17]. prostate
[Les15-30, NKW+19, TF19]. Protease [SBS+18, RDN+19, WLJ18, RFG19]. proteases
[BMC15, CC19]. protosomal [CYT+18, SPGB+17, UOT+16]. proteosome
[PAM+16, SSSR18, Sho18d]. protect
[AMT+15, BCM+19, CMTH+15, VZFG+18]. Protecting [IO18, OR17]. protection
[JJW17, RS19, Sho15-64]. protects [BMM+19, DWB+17, HSN+16, LUC+15, NLS+18, OPP+18, Sho16b, Sho16-27, WW16]. Protein
[BSL+15, BBK16, GNM16, HPB19, Nil19, RM19, RSvW+15, SM18, SS18, SB19, AWS+16, AKD+17, BPH+18, BCH+17, Boh18, BSP16, BG18, BPW+17, Can16, CJ16, CMM+15, CMB+18, CNN+17, DMC+16, DWH+17b, DLZ+15, DLBMA+15, FdSR+17, FKG+19, FCLoS19, GPP+18, GY18, GDB+15, GLS+15, Gli17, HKG17, HZB+15, IdSCB+16, IZZ+18, IG15, ISK+15, JHF+15, KMK+17a, KMK+17b, KJTY19, KMC+19, KML+15, LPWK15, LHY+19, Les15z, Les16i, LMPG+15, LSJY15, LCZ+16, LLW+17, LGH+18, LHB+18, MDOS19, MLJ+16, Mok16, MFP+17, MWF+15, NCV+16,
Quinlan [O’D17e].

WDM+15, WWT18, WCL+18, WSP+18, WHS+19, WKM+15, XWZ+15, YTGA16, ZPT+15, ZLG+15, ZGZ+15, ZAAN17, dlFEvW+15. **Regulating** [ABPS17, FG15, Har16, FBX+15, GCJ+15, GCH15, HSB+19, HQW15, IGK+16, IYP+18, KGN+15, LLK+17, LTS17, MpnDN+17, RDH+19, SSV+18, SVD+15, WYHG17, WBNH18, XJG+17, YAHH15, ZTR+17]. **Regulation** [CYH+16, CHS+17, CAl+15, CAA+17, Far16, KSL+17, LEM17, MF16b, PC17, RMMS+17, TJMM+18, TM18, ZAT+19, ABP+19, AOL+18, AKD+17, BBS+17, Can17, CLLL17, DZL+15, DB15a, FAH+17, GCD+19, GP17, HBH+19, HH18, HDA+17, HZM+15, JNS+19, KQM+19, LAMACE+17, LJ16, LJS+16a, LJS+16b, LZH+18, MGA19, MC16, MSK+19, MSL16, NHH17, Nil19, NPQ+17, OBS+17, PTMP+15, PMP+17, SJJ+19, SXT16, Sch17b, STF18, SK16b, SK18a, SL19, YWW17, ZCL+15, vDMR+19, vGWC+18, CL17]. **regulator** [CJ17, EMB+15, FG15, FdAV+17, KB16, PPB+15, QYP+17, QPZ+17, SLW+18, SIO+16, SIBM17, VWM+18, VMP16, WPA+18, Dic17]. **regulators** [BCMM+19, CPP+18, KJC+15, MCGM15a, MCGM15b, WZC+15]. **regulatory** [DUL+19, JGCAC+15, POTZ15, RND+17, SM18, SBC+16a, SBC+16b, VZB19, WBL+15, WHB+18, ZLG+15]. **reinnervation** [SCP+15]. **reintroduction** [KdBKvdK15, SKVvdK15]. **rejection** [vHGD+15]. **related** [BCMM15, CD18, DMC+16, MGE+15, VAB+18, CKKG17]. **relates** [LLZ+19]. **relations** [BFPD19]. **relationships** [CID17, TKM16]. **relatives** [CZM+18]. **relax** [GF16]. **relaxation** [GBD+18]. **relaxes** [EW17]. **RELCH/KIAA1468** [SiYM+18]. **release** [BLZ+15, BZG+17, CZZ+15, DNMB16, Das17, GSD+15, MWSM18, MWSM19, MPW+19, MG17, Nie19, NLH+19, PNE+19, PMW18, Rab17, SPD+17, SZSS18, SZL+16, SK18b, XJG+17, YWW17]. **released** [BJB+18]. **releasing** [KMJ+18]. **reliant** [ZCH+18]. **relies** [FFG+18]. **relieves** [Sho15-42]. **relieving** [LM15]. **relocalize** [IBFDB18]. **remain** [LJ17a]. **remnant** [Ott16]. **remodel** [LLAC18a, LLAC18b]. **remodeled** [CO19]. **remodeler** [GCA+17]. **Remodeling** [CAP+16, BPH+15, CRPSC+19, GCW+16, HHF15G15, KFAMR17, MEE+17, OMKM16, PLH18, SJJ+19, Sho16p, VCD+15, WW16, Lac19]. **remodels** [AKTR18, BMW+18]. **removal** [PhHS+16, PM15]. **remove** [MGO+18, YNN18]. **remyelination** [Sho15-72]. **renal** [DSH+18, LAMACE+17, SQJ+15]. **renewal** [TZC+15]. **reorganization** [CYT+18, DTM+16, LZD+16]. **reorientation** [BP19a, BP19b, LNS+19, LNL+18]. **Repair** [HSK+19, AWL18, ABGG16, BLL15, Can19, CPP+18, CR18, DQB+16, EEE+16, EMRS+18, GCZ+19, GCA+17, GRB19, GCW+16, HHF15G15, LWZ+18, LS16, LCD+17, LPHH16, Lov18, MCGC+15, MpnDN+17, NVP17, NIN+19, PLG+15, PMHB17, Pri17, QSZ+17a, QSZ+17b, SG19, Sho15-36, Sho17f, SOP+16, SJ16, XZ+18, YGM+17, vV17a]. **repairs** [RS16]. **Replicate** [Gom17]. **replicates** [LBG+17]. **replicating** [CST+16]. **Replication** [OO18, AWL18, BG18, BHS18, BCMM+19, CQB+19, CRS+17,
Col18, DKS15, Ger15, GRB19, HSN+16, Hyr15, Les16g, Lov18, MLJ+16, MN17, PST18, RS19, RLS18a, RLS18b, SPH+19, SD16b, TBL+15, UDH+16, WSP+18, XRH+18a, XRH+18b, YGMR+17, ZDM+15.

Retrogr...
IZZ+18, Sho15-51, LDR+19. secretases [CKS+15]. Secreted [HGD+15].
[JJB+19, MCS+15, PA19]. **Shaping**

[SBM17, JHC+16, LDG18, O’D17a, O’D17d]. share [Sho15-54]. **Sharma**

[Inf19a]. **Sharon** [Sed16e]. **She1** [ZAT+17]. shear

[BLO+16, FDR+16, Nic16]. **shed** [FD18]. **Shedding**

[HYC16, SAK+18, LFT+16]. **sheds** [RMOG17]. **sheet**

[FGR+18, OSL+19, WS18]. **shell** [Ver18]. shift [WHL17]. shifting

[CKM+16, May15, Pow15f]. **Shiga** [SIBM17]. **SHIP2** [RHC+16]. shock

[AB18, Can17]. **Shootin1** [KBT+15]. **short** [Les16e, McE19, vS15]. show

[JhZbYmp15, Sho15-69, SKO+15, ZZMC+15]. **showcases** [Les16a]. shows

[Les15d, Sho15-62]. **shrink** [Les15k, Sho16a]. **Shruff** [Jor16f]. **Shutting**

[vV17a]. **shuttle** [HR17]. side [BYMS+19, BBHBFS18, Jor16e, Sho18f]. **sides** [Sho15b]. **Sigma1** [SBP+16]. **signal**

[AS17, CHB+16, GGA+17, KBT+15, KDV+15, Sho17h, Sho17i, SCL+19]. signaling

[AUTM16, APHH+19, ASM+15, BJB+18, BBSA+16, BSP16, BDZ+15, CD18, CS16a, CAKL16, CIS+17, CNRR+17, CRA+19, CJ17, CAA+17, CKKG17, DGS+18, DLT+18, ED17, EPF16, FVF+16, FG16, FC19, FLS+16, GPAA+18, GKK16a, GKK16b, GDB+15, GP17, GLSS+15b, GLSS+15a, GCGAS+18, GKK16, GPD+19, GYS18, GKC+17, GKGK+18, HBM+19, HPW+17, HHS+16, HB18, IdSCB+16, JJJ+15, JRR+16, KDA+17, KG15, KZW+18, KBT+19, KNL+17, KLS+19, KNQ+19, LK+17, LDU+16, LR18, LFT+16, LKM+15, MBT16, MPW+19, MZN+17, OKN+16, OW+19, PDZ+18, PLD+15, RP18W17, RC15, SBS+18, Sch19, Sch17b, SSV+18, Sed15x, SD19, Sho15y, Sho17b, SL19, SYK+17, THA+16, TCWM18, TSJ+15, VB1+18a, VB1+18b, WBNH18, WV18a, WBL+15, XMJ+19, XG+17, YY+15, YPY+15, YSR+18, ZLG+15, dFEvW+15, vDM+19, vVFM+17, ODH19]. **signals** [CF15, CSTM+17, CPB+16, KMJ+18, LZC+15].

signature [SGB+17]. **signatures** [PCK+17]. silence [RMR17]. **Silencing**

[CHZ+17, LPRW17, LLS+16]. **Silva** [O’D19e]. **Simple** [TRM+16]. **simplicity** [TKG+19].

**Single**

[DCP+19, NHA+19, NiG+18, BYMS+19, ISL+18, PBS+16, PH16, SPJ+15, SLD+15, TBK+16, VB1+18a, VB1+18b]. **single-headed** [TBK+16].

**Single-molecule** [NiG+18, SPJ+15]. **single-particle** [SLD+15].

**singularity** [BDZ+15]. **SipA** [SKL+18]. **Sir4** [LPRW17]. **SIRF**

[RLS18a, RLS18b]. **SIRFing** [BG18]. **siRNA** [MLJ+16, SIBM17]. **SIRP**

[LBV+17]. **sisRNAs** [Sho15-52]. **sister** [SNB+18, TH18]. **sisters** [Lov18].

**site** [CGY+19, NP15, PAM+16, SSM+18, SZ17a]. **sites** [BDK+18, CVL+19, CR18, GSRG+18, GCW+16, GBM+15, KHA+18, KBJ16, KLHC+18, LFK+17b, MKS17, MSLK+18, MYN+17, NC18, OLL+17, PMHB17, PHA+17, RBP+17, SA19, SKZ+18a, VMA+15, VMA+15, VMA+15, ZNR+18].

situ [NDC+19, RLS18a, RLS18b]. **size**

[AOL+18, AWS+18, GM18, LDM15, LK17, PCP17, SWS+19].

**size-dependent** [AOL+18]. **Ska** [ACRM17, RGM+16]. **SKAP** [KNPC16].

**skeletal** [LNH+15, PLG+15, SAF+19, SFZ+17]. **Ski2** [VLZ15]. **Ski2-family** [VLZ15]. **skin** [CAI+15, FC15, Fuc15, Sho15-64, WZW+15]. **SLAMF1**
[ABF+16, BNKB15, KD17b, Sho15-58, ZCH+18, AvdH16, APHH+19, BBS+17, BTV16, BCS+17, BS17b, CHS+17, CSC+15, CO19, FFATC15, FTDc17, FC19, FBX+15, GHS16a, GHS16b, HAPC+19, HK15, IBFDB18, IWM+16, IG15, KNPC16, KY15, LSGM18, LDG+15, ML15a, MSLK+18, MGW18, iQLN+19, NHCB15, NDC+19, ORS+15, PUTM15, PCF+19, PMRM17, PDZ18, PCP17, RO18, RND+17, SLW+18, SFG+17, SPWM15, Sho15h, Sho15u, Sho15-59, SHO+18g, TWD+17, UOT+16, VGY+17, WGI16, Woo18, YLIW+15, YAH115, YTT15, ZLZD16, ZAT+17]. spindle-centering

[ZCH+18]. Spindle-E [ABF+16]. spindles

[Das17, GSC+16, Sho15w, Sho18c]. Spindly

[Sho15-60, GPS+17, MKA+17, MWF+15]. spine [WQD+18]. spines

[BBL+15, GSS+17, LMR+17, LIZD+16, LSS+15, Sho17a]. spindly

[ML+16], spinogenesis [RKK+18]. splice [Sed15]. spliced [RSC+19]. splicing

[MOCGD+17, OOT+18, PPB+15, RYS+15, VLZ15]. sporation [Sho16-32]. sporation

[BP19a, BP19b, Sed15]. spots

[BP19a, BP19b, HHT+16, Sho15-70]. Spotting [Pow15]. Spots

[SPL2] [NLS+18]. spread [Sho15-61, VZ17]. spreading [BVR+17, FVFI+16]. sprouting

[NWD+19]. spurs [Sho16-32]. SQST1 [WCY+16a, WCY+16b]. squeeze [LW16a]. Squeezing [SR17a]. SR [BMS+17, HR17]. Src [BBD+18, KG15, TQA+19, ANM+19, CEM+15, DPT+18, HHS+16, Sho18f, WWY+18]. Src-

[GBD+18]. Src-mediated [ANM+19, WWY+18]. Srf

[RHPH+18, FBBRCA+18]. SRP [FCLoS19]. SRP-receptor [FCLoS19].

[AGL+15, BRH+16, BRY+19, Can19, CQB+19, EFM17, HPB19, HSN+16, KBT+19, KGN+15, LLL+18, MGT+19, MHS+15, PBG+15, SSV+18, SFA+19, TT19, WGEH+18, XSJ18, YSM+17, ZAT+17]. Stabilization

[QYC+17, BSL+15, DBC+15, LNS+19, PSL+17, SID+18]. stabilize

[BG1+16, DSvNA+15a, DSvNA+15b, KMC+19, RHC+16]. stabilizes [ASZ+18, ALY+17, DLZ+15, EG19, GBK+17, GBM+15, JKD+19, NIS+16, SMA+19]. stabilizing [NKP+15]. Stable

[POTZ15, BGH18, GCL+15, KMBO+15, LCP+15]. stably

[CSF+17, CSF+18]. stacks [Les15]. stage [ITN+17, WGI16]. stages

[NDL17]. stalled [Les15, RS19]. Stardust [DK17, PMRMS17]. starfish

[BPSK+16, Ver16]. start [Les15]. starting [Sch15]. Starvation

[MOS+18, MTGG18, NPU+16, SvsZ+16, VTG+16, VLvdKR18]. Starving

[Les15-27]. STAT3 [RMMS+17]. state

[BMS+17, HLW+15, HAPC+19, SRI+19]. states

[JP+16, RRR+17, VRK+17, WVM+18]. static [Nel17]. stationary

[GSKL+18]. status [MP18]. stay [FV17, Sho15-32, Sho16a]. Staying [SA19].

[Sto5] [vDMR+19]. STED [SBM+19]. steering [WRH+16]. steers [Les15o]. Stem

[KF18, ASPY+16, BSK+19, CNC+18, CSG+15, CEM+15, CM18, DSC+18, FG15, GBHR15, GCC+18, KPEJ17, LLK+17, Les15y, LWF+15, PLG+15,
PA19, RMB⁺18, SZF⁺15, Sed15e, Sho15-62, Sho16c, Sho17g, SR17a, SCP⁺17, TGG⁺17, TST⁺17, TSJ⁺15, UGH⁺16, VY18, VZFG⁺18, WCY⁺16a, WCY⁺16b, WHB⁺18, XTS⁺15, YLND⁺16, ZGDS⁺16, THG19.


summits [O'D18f]. SUMO [RFG19, Sho15-63]. SUMO/protease [RFG19].

Taking [FB15, LS16, Pfe16, Sho15q, Sho15-47, Sho16-37]. TALE [Les16g, SA19, Jor16f, Pow15i, Pow15j]. TALI [SNOBM16]. talin [GLC+19, GYS18, KOV+16a, KOV+16b]. talk
tango [Sho15q, Pfe16]. TANGO1 [Gli17, MKS17, RBP+17, SNOBM16, LFK+17b]. Tapping [DK16, Yud19].
target
[AMS+17, CMM+15, KV+17, MTN+16, SPMM+17, SAK+18, SCP+17]. targeted [HHT+16, NDL17, OSR+15]. targeting
[BHS+16, DMS+15, HPB19, IWM+16, KJC+15, SKZ+18b, WLJ16]. TAT1 [FFBRC+18]. TATA [PLH18]. Tatsushi [O’D18f]. Tau
[QYC+17, SID+18, VXF+15]. Tau-dependent [QYC+17]. taxol [MHA+16]. taxol-treated [MHA+16]. TAZ [MCD+19, PGRY+19, GCC+18, NW+19].
tectum [CED+15, Les15]. telangiectasia [BLO+16]. tell [Sho15z]. telomerase [OLL+17]. Telomere
[MB17b, AGGSF+16, CG17, DKS15, Ger15, LSJY15, RSG+15]. telomere-associated [LSJY15]. Telomere-driven [MB17b].
telomere-targeting [MB17b]. Telomeres
[FFÁTC15, Sho16-33, DKS15, Sho15-73]. template [cLNF+16]. temporal
[AS17, GGA+17, HPB19, WLC+17, YVIMS18]. temporally [BMP+18].
tensile [KJZ+19, MRO+15]. Tensin [DN17, GLJ+17]. tensin-dependent
[GLJ+17]. tension [CAP+16, DSvNA+15a, DSvNA+15b, KS17, KD17b, KOV+16a, KOV+16b, LVE+15, LDMW+15, MHA+16, PMG+17, RMB+19, Sho15-65, TCD+15, TNK18]. tensions [BHS18]. tent [CHC+18, PH+18].
term [FTAB+15]. Terminal
[vHG+15, DAG+15, GDV19, LHZ+18, OBY+15, SCL+16, NL16, SER+15]. terminals [FSF+15, KG19]. termination [IKK+18]. terminus [MRWM18].
test [Sho16q, Sho17a, SLH17]. testosterone [GLL+18a]. tether
[FR16, HCC+17, Lac19, LCTP17, SJ+19, SES+19]. tethered [PLD17].
tethering [HZB+15, MGW18, NO19, QYY+16, TCP+15, XLM+18]. tethers
[AKTR18, CWT+19]. tetramer [FKO+18]. tetraspanin [OPP+18].
tetraspanins [DCO+12, DCO+16]. TFE [NWY+15]. TGF
[DKA+16, YVZ+15]. TGF- [DKA+16]. TGN [DOA+17, VMR+19]. their
therapeutic [MG18]. therapies [MB17b, TG19]. therapy [ASPY+16].
There [Sho17]. thereby [WEQ+15]. things [Jor16d, O’D19d, Sed16d].
Thinking [MC15, She15, SB17]. thiol [RGOS⁺16]. thiol-based [RGOS⁺16].
Thirty [HK15]. Thomas [Sed15w]. Thoru [Pow15k]. thought [Yel18].
Three [FAH⁺17, Sho16-34, VLP⁺15, JhZbYmP15, MSW⁺07, MSW⁺17, MG16, Sho17i, SB17]. three-alarm [Sho17i]. three-step
[MSW⁺07, MSW⁺17]. Three-tier [FAH⁺17]. threshold
[WXC⁺18, XPZ⁺19]. thrombopoiesis [NNK⁺15]. Thrombospondin
[RKK⁺18]. throughout [MGA19]. throughout [BCG⁺19, Pow16d].
Thy [FSB⁺15, Sed15x]. Thy-1 [FSB⁺15, Sed15x]. thyself [Sho15g].
Tiam [GKK16a, GKK16b]. ticket [Sho15i]. tier [FAH⁺17]. tight
[ONT⁺19, SOI⁺18, SLM⁺15, TE15]. tiki [LPHH16]. Tilted [FGR⁺18].
TIM23 [RPMC⁺16]. Time [CSA19, WHB⁺18, BPW15, FJ17, Gom17, Jor16c, Les16f, MSvO17, O’D18a, SP⁺15, SHC⁺18]. Time-resolved
[CSA19, WHB⁺18]. timeline [Sho15-66]. timely
[ABPS17, DOH⁺17, SOP⁺16]. timer [BMP⁺18, MF16a]. timing
[MN17, OO18, PST⁺18]. TIMP [SAF⁺19]. tip
[Les15m, LHB⁺18, NDRJ15, YVM18]. Tipping [AvdH16]. tips
[MOM⁺18, Sed15t, Sho15-71, Sho16i, THA⁺16]. Tissue [HF15, LLS⁺16, CPP⁺18, Jan18, JNS⁺19, KS19, KTM19, KQM⁺19, LDP⁺15, O’D16b, PLG⁺15, SBM17, Sho17l, SBC⁺16a, SBC⁺16b, SLG⁺18, TY16, YEM⁺19].
tissues [FBPN⁺18, Nel17]. Titration [CRC⁺15]. Tks5 [CLO⁺19].
Tkv [LWF⁺15]. TLR3 [HCS⁺18]. TLR4 [HS16, SQC⁺16, YSR⁺18].
TLR4-mediated [YSR⁺18]. TLR7 [HCS⁺18]. TLR8 [HCS⁺18].
TMEM231 [RDO⁺15, Sho15-67]. TMEM41B [MHI⁺18]. TMX1
[RGOS⁺16]. TNT [SS19]. TNT-like [SS19]. TOG [BS17b]. TOG-tubulin
[BS17b]. together
[MB17a, O’D18c, O’D18g, Sed15k, Sed15w, Sho18b, TH18]. tolerance
[RMS⁺18, YGMR⁺17]. tolerogenic [VRK⁺17]. Toll [MNLB16, OG16].
Toll-like [OG16]. Toll-tally [OG16]. Tolls [FAH⁺17]. TOM [WEQ⁺15].
Tom22 [WLJ18]. Tom70 [BHH⁺18]. tool [ISK⁺15, LLZ⁺19]. toolbox
[PBG18]. tools [BOL17, WLC⁺17]. tooth [AUTM16, SK16a]. TOPBP1
[LS16, LCD⁺17, MWW⁺16, SG17, PKN⁺15]. TOPII [MGSO⁺18]. Topo
[Sho16-36]. topoisomerase [ABGG16, EJK⁺16, YTGA16, LRS⁺17].
topology [GRU18, MRWM18, SLG⁺18]. TORC1
[MP17a, MYN⁺17, vdVF⁺17]. TORC1-independent [MP17a]. TORC2
[LT19b, MYN⁺17, RBM⁺19, VMP⁺16]. TORC2-mediated [VMP⁺16].
touchy [Les15p]. touchy-feely [Les15p]. toxicity [MCH⁺18].
toxin [LH19, SIBM17]. Toxoplasma [RNP⁺17]. TPX2
[AATP17, BCMM⁺19, Can19, FBX⁺15]. TPXL [MSK⁺18]. TPXL-1
[MSK⁺18]. TR [GX16]. track [Ava18, EFM17, Sho15-53]. Tracking
[DB15b, Sho16-35, JPD⁺16, SP⁺15, PH16, Pow15h]. traction
[DPGS⁺18, JhZbYmP15]. traffic
[DS16b, HHT⁺16, Inf19a, Jor16e, MAJ⁺17, MP17b, WDM⁺15, WHS⁺19]. Trafficking
[dlRHM⁺18, BhHS⁺17, BKH⁺15, BBC⁺16, CWI⁺19, CJ17,
DCO+12, DCO+16, DLBMA+15, EPF16, FWL+17, Far16, FCB+09, FCB+19, GLS+15, GSS+17, IM16, MRGWB+16, MSCS19, MFP17, MF16b, ODH19, SSC+19, SIBM17, Sor17, ZsdD+15, ZWS+16]. trail

TRAMM/ [MHSD+15]. tranquilizes [Sho15y]. trans
[CBM+16, DsvNA+15a, DsvNA+15b, GKK16a, GKK16b, Sed15p].
trans-endocytosis [GKK16a, GKK16b]. trans-Golgi [CBM+16, Sed15p].
trans-interaction [DsvNA+15a, DsvNA+15b]. Transcription
[EMRS+18, BMP+18, BGH18, Cas17a, CDF+18, DAG+15, Ger18, HHBG17, LT18, NWFY15, PBC+15, RHCS+16, SPK+18, Sed16b, TSB+18, UD+16].
Transcriptional [OBY+15, SK18a, VRK+17, Bob17, CMM+15, CIK+17, DMG+19, FBBRCA+18, GVCAGS+18, HCS+18, KF18, QJP+17, SIO+16, SXT16, XMJ+19]. transcriptome [LJ16]. transcriptomics [WHB+18].

transcytosis [NDL17]. transducer [ITN+17]. transduces [VCD+15].
Transducing [CF15]. transduction [KBT+15]. transfer
[AFO+16, CPCtr+15, GY18, KTK+18]. transferrin [DNMB16].
transforming [KKD+16]. Transient [LXR+15]. transit
[BGH18, WWW+18]. Transition
[VPD+16, DCF+17, LW+19, RDO+15, SXT16, SSPD15, YNN18].
translating [KP18, O’D19e]. Translation
[PBL+19, PBS+16, PH16, SPMM+17, SG18a, SG18b, VLP+15].
Translational [NPÖ+17, Inf18c, O’D19a, SL19]. translationally
[SEN+15]. translesion [Sho15-36, TSFP+15]. translocase
[CS+17, RDN+19]. translocates [PLH18]. translocation
[CCQ+18, GDV19, IkrRM16, SSL+17, TYD+15, WYo17]. translocon
[KDV+15, SCG17]. Transmembrane [GSM+15, GKG+18, SLG+18].
transmigration [NLH+19]. Transmission
[SLG+18, KTM19, KOV+16a, KOV+16b, PKN+15]. transmitted [FSF+15].
transport [AGB+19, BMF+18, BCH+17, CPBG19, CDT+19, CNN+17, CGPB17, CZL+15, CHI+15, CCV+19, DOA+17, Ddar+16, GHD+17, GYK+17, GWF17, JERL+15, JNW15, KKC+19, KHRL17, KLHC+18, KOK+19, KJON+17, LCTP17, LE16, LLS+18, LDG+15, Mes16, MHA+19, MWT+16, NiYT+16, NH17, OI18b, OOT+18, RM19, RFG19, SIO+16, SD19, SMK+18, Sho15-35, SDHC17, VGB+17, VXV+15, XTT+18, YTH+17, YDM+18, YWW17, YSM+17, ZYL+16]. transported
[ADBST+15]. transporter [GWZ+19, HDA+17, MST+15]. transporters
[MYN+17]. transports [SS19, TBK+16, VYB+19]. trap [SAO+17].
TRAPP [RGMM18]. TrappC12 [MHSD+15]. TRAPP II [TF16].
TRAPPII [TF18]. trapping [GHC+17]. traps [WWT18]. trash
[Blu15b, VR18]. travels [SS19]. TrCP [XWZ+15]. Tre1 [LL17, TCWM18].
treated [MHA+16]. treatment [BS18]. treatments [ZDM+15]. TREX
[EMRS+18]. TREX-2 [EMRS+18]. triage [Sed15n]. tricellular [SLM+15].
trichoplein [IGK+16]. TRIF [YSR+18]. TRIF-dependent [YSR+18].

Uncoordinated [YYM+18]. uncoupled [LJ17a]. underlies
[DKM+15, SPWM15, YYM+18]. understanding
[Inf19b, Jor16a, LS18, RS19, Mar17]. Unearthing [Pow15g]. unequal
[LLS+16]. unexpected [Mok16]. unfolding [Sed16e]. Unidirectional
[OKN+16]. Unipotent [WRGB+15]. Unique [PCK+17, VAB+18].
universally [DBS18]. unlicensed [CNC+18]. unpredictability [LH15].
up-regulation [ZCL+15]. uPARAP [JNS+19]. uPARAP-mediated
[JNS+19]. upon
[KdBKvdK15, LCP+15, LBV+17, LMC+18, MTM+17, SSRG18]. UPR
[ITN+17]. upregulated [CHH+15]. ups [ZZ16]. Ups2 [AFO+16, MWT+16].
upstream [FG15]. uptake [CJS+18, GLL+18a, JNS+19, LLZ+19]. use
[Ava18, PLD17, Sho15f]. Usher [SPE+17a]. Using [Les16i, AHS+18,
BCG+19, CZV+18, ISL+18, Juh16, SERP16, TCWM18, BA18]. UsnRNP
[PMP+17]. USP10 [KPA+16]. Usp16 [ZGZ+15]. USP28
[LDU+16, MAK+16]. USP30 [RDH+19]. USP9X [KMC+19]. utilizing
[AKTR18].

\( v \) [ZT15, DB15a, TBK+16, YWW17, WHB+18]. \( v\text{-ATPase} \) [WHB+18].
\( v\text{-ATPase/WHB+18}. \) vaccinia [PMW18]. vacuole
[AKTR18, BPL+18, JJJ17, PHA+17, SE19, Sho15-32, SE18, MST+15].
vacuole/lysosome [JJ17, SE18]. vacuoles [GRU18]. vagaries [Pow16a].
Validating [JJ19]. valve [GGF+19, WBL+15]. VAMP7
[DDAR+16, VJ15]. VAMP8 [MPH+15]. VAMP8-dependent
[MPH+15]. VAPB [CCH+17]. VAPs [HCC+17]. variant [RSC+19].
variants [OOT+18]. varicosity [GJW+17]. varying [Ol18b]. Vascular
[YGW+17, LWZ+18, VAKB+18]. vasculature [Sho17e]. VASP [LWZ+18].
Vav3 [HNF+18]. Vav3-induced [HNF+18]. VCAM [LAMACE+17].
VCAM-1 [LAMACE+17]. VDAC2 [iHMM+17]. VE
[CBH+15, DsvNA+15a, DsvNA+15b, GDB+15, JKD+19, Sho15v].
VE-cadherin
[CBH+15, DsvNA+15a, DsvNA+15b, GDB+15, JKD+19, Sho15v]. VE-PTP
[JKD+19]. VEGF [Sho15-71]. VEGFR2 [CBH+15]. VEGFR3 [CBH+15].
velocity [AGB+19]. Ventura [Cas17b]. versatile [GSC+16]. versus
[DS16a, ST16a]. vertebrate [HKT+17, SSdLA+15]. vertebrates [OTG+18].
vesicle [BLPV+17, JHC+16, Juh16, KCB+16, KMK+17a, KMK+17b,
Sho15-35, TF16, UBBSM15]. vesicles
[BBZ+17, DB15b, JPD+16, KMJ+18, MLMF16, NN17, RM19, RNP+17,
SES+19, SZSS18, SKN19, SCP+17, SW18, ZIM+17]. vessels
[FG16, Pow16a, TCWM18]. VHL [LAMACE+17]. VI [RHH+18]. via
[ASZ+18, ASM+15, BKH+15, BJ18, CBAP+17, CAKL16, CMNB+15,
CRS+17, CCY+19, DWH+17a, DDAR+16, DCB+15, FBBRCA+18, FRP+17,
Gcz+19, GCC+17, HKH16, JKD+19, KG15, KSM+18, KLS+19, KD17b,
KJF+18, LAMACE+17, LSMG18, LHY+19, LMPG+15, LJ17b, LZD+16,
LZH+18, LSS+15, MFVS18, MBF17, MBC+19, NHA+19, OMKM16, PLH18, PLS+15, PAC+15, PMRMS17, PKC+16, PBL+16, RKK+18, SS19, SOII18, SKZ+18a, SK18b, SYK+17, TTU+17, UBBSM15, WWZ+18, WSP+18, XJ18, YVM18, YKO+16, vDMR+19. viability [BRY+19, SFZ+17].


REFERENCES

[Sed15u]. works [Sho15b]. wound
[CPP+18, HLHFG15, LLC+17, MCGM15a, MCGM15b, NVP17, ZPT+15]. wounds [TCWM18]. Wrapping [Boh18].

X [CAI+15, OSW+17, dIFeW+15]. XBP1 [TSK+18, TSK+19]. Xenopus [GHS16a, GHS16b, GSC+16, KZW+18, MH15, RBR19]. Xiaochen [Inf19b]. XLF [CQB+19, GRB19]. XPA [CR18]. Xpo7 [APK+18]. Xrpl [MCH+18].


References


Andress:2016:SCB


Aparicio:2016:MCB


Adriaans:2019:PPD


Alleva:2017:RCM


Aulas:2015:GPS

Anaïs Aulas, Guillaume Caron, Christos G. Gkogkas, Nguyen-Vi Mohamed, Laurie Destroismaisons, Nahum Sonenberg, Nicole Leclerc, J. Alex Parker, and Christine Vande Velde. G3BP1 promotes stress-induced RNA granule interactions to preserve polyadenylated mRNA. *Journal of Cell Biology*, 209


Aaltonen:2016:MPT


Asai:2019:ABK


Agrawal:2016:DRI


Atkins:2019:FAK

Melody Atkins, Laïla Gasmi, Valérie Bercier, Céline Revenu, Filippo Del Bene, Jamilé Hazan, and Coralie Fassier. FIGNL1 associates with KIF1Bβ and BICD1 to restrict dynein transport velocity during axon navigation. *Journal of Cell Biology*, 218(10):3290–??, October 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/10/3290.
Aix:2016:PTD


Aminí:2015:CEA


Awadia:2019:SFC


Amano:2015:KIS


Adikes:2018:CMD

Aird:2016:HOC


Azad:2018:PDE


Avenarius:2017:HCP


Arasaki:2018:LRP

REFERENCES


[ALY+17] Ming-Xin An, Si Li, Han-Bing Yao, Chao Li, Jia-Mei Wang, Jia Sun, Xin-Yu Li, Xiao-Na Meng, and Hua-Qin Wang. BAG3 directly stabilizes Hexokinase 2 mRNA and promotes aerobic glycolysis in pancreatic cancer cells. *Journal of Cell Biology*, 216(12):4091–??, December 2017. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/12/4091.


REFERENCES

Allard:2018:CSD


Alfonso-Perez:2019:MDR


Aksu:2018:XBS


Ando:2017:NDP


Aitchison:2015:IC

Aweida:2018:GPC


Atay:2017:STS


Artym:2015:DF


Aratyn-Schaus:2016:CPS


Agarwal:2018:CSK

REFERENCES

3446–??, October 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/10/3446.

Aksu:2019:SBN


Ahtiainen:2016:EES


Avasthi:2018:CMM


Akhmanova:2016:TSR


Amarh:2018:DRM


Brown:2017:IPR


Bruno:2016:SRE


Broders-Bondon:2018:MTP


Bersuker:2016:PMS


Biname:2016:CAM

REFERENCES


REFERENCES


REFERENCES

Bean:2018:COS

Bentley:2015:NAR

Brace:2019:CSC

Bruurs:2015:AMS

Beaudoin:2016:MCP
Gerard M. J. Beaudoin. Mosaic cellular patterning in the nose: Adhesion molecules give their two scents. *Journal of Cell Bi-

REFERENCES


Bayless:2016:ALP


Bezanilla:2015:CDV


Bond:2015:LSG


Backes:2018:TEM


Burke:2017:BVF

Badgandi:2017:TFP


Balachandran:2016:ULC


Brayson:2018:MTM


Bonner:2019:EAB


Bernkopf:2018:PRD


Yi Bao, Carola Ledderose, Amelie F. Graf, Bianca Brix, Theresa Birsak, Albert Lee, Jingping Zhang, and Wolfgang G. Junger. mTOR and differential activation of mitochondria

**Badrinarayanan:2015:RPR**


**Baeyens:2016:DFS**


**Blow:2019:DOL**


**Buono:2017:EMV**


**Blum:2015:MSL**


REFERENCES


Belvindrah:2017:MTT


Bobola:2017:DBT


Bohnert:2018:WFS


Bugaj:2017:ICP


Benoit:2019:CMR

REFERENCES

Benoit:2019:MRB


Bonello:2019:SMS


Bassler:2015:NAF


Beati:2018:AJA


Bas:2018:RRY

Levent Bas, Daniel Papinski, Mariya Licheva, Raffaela Torggler, Sabrina Rohringer, Martina Schuschnig, and Clau-


REFERENCES


[Bowes:2019:CDR] Charnese Bowes, Michael Redd, Malika Yousfi, Muriel Tauzin, Emi Murayama, and Philippe Herbomel. Coronin 1A deple-

Bezbradica:2017:ZES


Byrnes:2017:TTB


Bansal:2018:GMC


Belair:2019:REN


Bencsik:2015:PKD


Ralph T. Böttcher, Maik Veelders, Pascaline Rombaut, Jan Faix, Marina Theodosiou, Theresa E. Stradal, Klemens Rottner, Roy Zent, Franz Herzog, and Reinhard Fässler. Kindlin-2 recruits paxillin and Arp2/3 to promote membrane protrusions
<table>
<thead>
<tr>
<th>Reference</th>
<th>Authors</th>
<th>Title</th>
<th>Journal</th>
<th>Volume/Issue</th>
<th>Month</th>
<th>Year</th>
<th>DOI</th>
<th>URL</th>
</tr>
</thead>
<tbody>
<tr>
<td>[BYMS+19]</td>
<td>Rakefet Ben-Yishay, Amir Mor, Amit Shraga, Asaf Ashkenazy-Titelman, Noa Kinor, Avital Schwed-Gross, Avi Jacob, Noga Kozer, Pramod Kumar, Yuval Garini, and Yaron Shav-Tal</td>
<td>Imaging within single NPCs reveals NXF1’s role in mRNA export on the cytoplasmic side of the pore</td>
<td>Journal of Cell Biology</td>
<td>218(9)</td>
<td>September</td>
<td>2019</td>
<td>10.1083/jcb.201903101</td>
<td><a href="http://jcb.rupress.org/content/218/9/2962">http://jcb.rupress.org/content/218/9/2962</a></td>
</tr>
<tr>
<td>[BZG+17]</td>
<td>Joseph J. Bruckner, Hong Zhan, Scott J. Gratz, Monica Rao, Fiona Ukken, Gregory Zilberg, and Kate M. O’Connor-Giles</td>
<td>Fife organizes synaptic vesicles and calcium channels for high-probability neurotransmitter release</td>
<td>Journal of Cell Biology</td>
<td>216(1)</td>
<td>January</td>
<td>2017</td>
<td>10.1083/jcb.201609130</td>
<td><a href="http://jcb.rupress.org/content/216/1/231">http://jcb.rupress.org/content/216/1/231</a></td>
</tr>
<tr>
<td>[CAA+17]</td>
<td>Helen Court, Ian M. Ahearn, Marc Amoyel, Erika A. Bach, and Mark R. Philips</td>
<td>Regulation of NOTCH signaling by RAB7 and RAB8 requires carboxyl methylation by ICMT</td>
<td>Journal of Cell Biology</td>
<td>216(12)</td>
<td>December</td>
<td>2017</td>
<td>10.1083/jcb.201707011</td>
<td><a href="http://jcb.rupress.org/content/216/12/4165">http://jcb.rupress.org/content/216/12/4165</a></td>
</tr>
<tr>
<td>[CAI+15]</td>
<td>Byung Min Chung, Artem Arutyunov, Erika Ilagan, Nu Yao, Marsha Wills-Karp, and Pierre A. Coulombe</td>
<td>Regulation of...</td>
<td>...</td>
<td>...</td>
<td>...</td>
<td>...</td>
<td>...</td>
<td>...</td>
</tr>
</tbody>
</table>


REFERENCES

CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/213/2/243.


F-actin flow drives affinity maturation and spatial organization 
of LFA-1 at the immunological synapse. Journal of Cell Biology, 208(4):475–??, February 2015. CODEN JCLBA3. ISSN 
rupress.org/content/216/9/2979.

[CBF+18] Ivan Nicola Colaluca, Andrea Basile, Lee Freiburger, Veronica 
D’Uva, Davide Disalvatore, Manuela Vecchi, Stefano Con 
falonieri, Daniela Tosoni, Valentina Cecatiello, Maria Grazia 
Malabarba, Chun-Jiu Yang, Masatsune Kainosho, Michael 
Sattler, Marina Mapelli, Salvatore Pece, and Pier Paolo Di 
Fiore. A Numb-Mdm2 fuzzy complex reveals an isoform-
pecific involvement of Numb in breast cancer. Journal of 
ISSN 0021-9525 (print), 1540-8140 (electronic). URL http:// 
jcb.rupress.org/content/217/2/745.

[CBH+15] Brian G. Coon, Nicolas Baeyens, Jinah Han, Madhusudhan Bu 
datha, Tyler D. Ross, Jennifer S. Fang, Sanguk Yun, Jeon-Leon 
Thomas, and Martin A. Schwartz. Intramembrane binding of 
VE-cadherin to VEGFR2 and VEGFR3 assembles the endo 
thelial mechanosensory complex. Journal of Cell Biology, 208 
(7):975–??, March 2015. CODEN JCLBA3. ISSN 0021-9525 
org/content/213/3/305.

[CBM+16] Alvaro H. Crevenna, Birgit Blank, Andreas Maiser, Derya 
Emin, Jens Prescher, Gisela Beck, Christine Kienzle, Kira 
Bartnik, Bianca Habermann, Mehrshad Pakdel, Heinrich Leon 
hardt, Don C. Lamb, and Julia von Blume. Secretory cargo 
sorting by Ca2+-dependent Cab45 oligomerization at the trans-
Golgi network. Journal of Cell Biology, 213(3):305–??, May 
2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 
(electronic). URL http://jcb.rupress.org/content/213/3/ 
305.
REFERENCES

Chang:2019:BPB


Costello:2017:AVM


Chen:2017:RCS


Chang:2018:EBR


Chow:2019:AAA


Cooper:2015:PCM


Chetty:2015:SIR


Cao:2015:TOS


Catlett:2016:DLG


Churikov:2017:NTA


Cruz-Garcia:2017:DMD


Cheat:2017:ATM


Chang:2017:RMS


Cuellar:2017:SRS


Cho:2017:MNP


Chau:2017:MCC

[CIK+17] Gia Cac Chau, Dong Uk Im, Tong Mook Kang, Jeong Mo Bae, Won Kim, Suhkneung Pyo, Eun-Yi Moon, and Sung Hee Um.
REFERENCES


**Capurro:2017:GPG**


**Casadio:2016:JFK**


**Colaco:2017:RMR**


**Chakrabarti:2018:IMA**


**Chen:2015:ROC**

[CKJ+15] Jieyan V. Chen, Ling-Rong Kao, Swadhin C. Jana, Elena Sivan-Loukianova, Susana Mendonça, Oscar A. Cabrera, Priyanka Singh, Clemens Cabernard, Daniel F. Eberl, Monica Bettencourt-Dias, and Timothy L. Megraw. Rootletin or-

**Cullati:2017:BSC**


**Capote:2016:OAA**


**Chen:2015:PFI**


**Chen:2016:AMD**

Cunningham:2019:NRS


Comrie:2015:DCC


Csizmadia:2018:MMD


Chiapparo:2016:MCS


Lin:2016:MTC

REFERENCES

Chuang:2019:TDE


Chipuk:2017:PSB


Chen:2016:CAJ


Castro:2018:DGR


Chang:2018:CGE


REFERENCES


Casler:2019:MDT


Carroll-Portillo:2015:MCD


Colombelli:2015:PRD


Carvalho:2018:OJN


Chen:2019:XHF

[CQB+19] Bo-Ruei Chen, Annabel Quinet, Andrea K. Byrum, Jessica Jackson, Matteo Berti, Saravanabhavan Thangavel, Andrea L. Bredemeyer, Issa Hindi, Nima Mosammaparast, Jessica K. Tyler, Alessandro Vindigni, and Barry P. Sleckman. XLF and
<table>
<thead>
<tr>
<th>Reference</th>
<th>Title</th>
<th>Journal Details</th>
<th>URL</th>
</tr>
</thead>
<tbody>
<tr>
<td>CRK+17</td>
<td>Hyo Kyun Chung, Dongryeol Ryu, Koon Soon Kim, Joon Young Chang, Yong Kyung Kim, Hyon-Seung Yi, Seul Gi Kang, Min Jeong Choi, Seong Eun Lee, Saet-Byel Jung, Min Jeong Ryu, Soung Jung Kim, Gi Ryang Kweon, Hail Kim, Jung Hwan Hwang, Chul-Ho Lee, Se-Jin Lee, Christopher E. Wall, Michael</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Title</td>
<td>Authors</td>
<td>Journal</td>
<td>Volume</td>
</tr>
<tr>
<td>-------</td>
<td>---------</td>
<td>---------</td>
<td>--------</td>
</tr>
<tr>
<td>Growth differentiation factor 15 is a myomitokine governing systemic energy homeostasis.</td>
<td>Downes, Ronald M. Evans, Johan Auwerx, and Minho Shong.</td>
<td><em>Journal of Cell Biology</em></td>
<td>216</td>
</tr>
</tbody>
</table>
REFERENCES


REFERENCES


REFERENCES


Sandip Dey, Chiranjit Biswas, and Jayati Sengupta. The universally conserved GTPase HflX is an RNA helicase that re-

**Donninger:2015:NRS**


**Dyson:2017:IRP**


**Donnelly:2017:RRB**


**Dornier:2012:TTR**

[DCO\textsuperscript{+16}] Emmanuel Dornier, Franck Coumailleau, Jean-François Ottavi, Julien Moretti, Claude Boucheix, Philippe Mauduit, François Schweisguth, and Eric Rubinstein. Correction: TspanC8 tetraspanins regulate ADAM10/Kuzbanian trafficking and promote Notch activation in flies and mammals. \textit{Journal of Cell Biology}, 213(4):495–??, May 2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL \url{http://jcb.rupress.org/content/213/4/495}. See [DCO\textsuperscript{+12}].


[DER\textsuperscript{+18}] Rachayata Dharmat, Aiden Eblimit, Michael A. Robichaux, Zhixian Zhang, Thanh-Minh T. Nguyen, Sung Yun Jung, Feng He, Antrix Jain, Yumei Li, Jun Qin, Paul Overbeek, Ronald

\url{http://jcb.rupress.org/content/214/3/293}.
REFERENCES


Shradha Das and Elisabeth Knust. Stardust, the Janus-faced partner of Crumbs. *Journal of Cell Biology*, 216(5):1219–??,


[Dwivedi:2019:DAH] Devashish Dwivedi, Amrita Kumari, Siddhi Rathi, Sivaram V. S. Mylavarapu, and Mahak Sharma. The dynein adaptor...


Derive:2015:BBD


delaRoche:2018:TCR


Drummond:2018:APC


Draheim:2015:CCI


DeLuca:2018:AKP

Keith F. DeLuca, Amanda Meppelink, Amanda J. Broad, Jeanne E. Mick, Olve B. Peersen, Sibel Pektas, Susanne M. A. Lens, and Jennifer G. DeLuca. Aurora A kinase phosphorylates Hec1 to regulate metaphase kinetochore–microtubule dy-


REFERENCES


REFERENCES


Deshpande:2016:BSM


Signore:2019:EME


David:2019:AAL


Davis-Roca:2017:CEO


Dantuma:2016:UVM

REFERENCES


REFERENCES


REFERENCES

ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/12/4299.


REFERENCES


REFERENCES


REFERENCES


**Efmova:2018:BAN**


**Ebner:2017:LMA**


**Elazar:2019:CIP**


**Elzeneini:2017:LLL**


**Ewers:2018:OSR**

REFERENCES


REFERENCES


REFERENCES

Franco:2019:ESC


Franca:2009:BNF


Franca:2019:CBN


Fu:2019:MRF


Felce:2018:NKS


kinesin–II with IFT–B–connecting tetramer is crucial for cil


[Ferron:2015:GVI]


[Forest:2018:ASB]


[FLG+19]
REFERENCES


[FML+17]
REFERENCES


REFERENCES


REFERENCES


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/208/2/143.

Garrett:2015:CBM


Grimsey:2015:UPA


Graham:2018:ECR


Gemble:2018:FFL


Goel:2019:HSA

REFERENCES


REFERENCES


Guo:2018:KRM


Gomez-Cavazos:2015:NGN


Gerber:2015:RCH


Gluszek:2015:MCP

REFERENCES

Gomis-Coloma:2018:CIH


Gracheva:2016:ZMR


Galino:2019:RCS


Green:2016:CBA


Giampietro:2015:ABP


REFERENCES

0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/10/2999.


REFERENCES

Alessandro Ori, Khanh Huy Bui, Jochen Baßler, Elisar Bar-
bar, Martin Beck, and Ed Hurt. Structural basis for assembly
and function of the Nup82 complex in the nuclear pore scaffold.
*Journal of Cell Biology*, 208(3):283–??, February 2015. CODEN
JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL
http://jcb.rupress.org/content/208/3/283.

Katharina Grikscheit, Tanja Frank, Ying Wang, and Robert
Grosse. Junctional actin assembly is mediated by Formin-like
2 downstream of Rac1. *Journal of Cell Biology*, 209(3):367–??,
May 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-
8140 (electronic). URL http://jcb.rupress.org/content/
209/3/367.

Ana Guerrero and Jesús Gil. HMGB2 holds the key to the
senescence-associated secretory phenotype. *Journal of Cell Bi-
ology*, 215(3):297–??, November 2016. CODEN JCLBA3. ISSN
rupress.org/content/215/3/297.

Brian R. Graziano, Delquin Gong, Karen E. Anderson, Anne
Pipathsouk, Anna R. Goldberg, and Orion D. Weiner. A mod-
ule for Rac temporal signal integration revealed with optogenet-
ics. *Journal of Cell Biology*, 216(8):2515–??, August 2017. CO-
DEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic).
URL http://jcb.rupress.org/content/216/8/2515.

Tullio Genova, Guillaume P. Grolez, Chiara Camillo, Michela
Bernardini, Alexandre Bokhobza, Elodie Richard, Marco
Scianna, Loic Lemonnier, Donatella Valdembri, Luca Munaron,
Mark R. Phillips, Virginie Mattot, Guido Serini, Natalia Pre-
varskaya, Dimitra Gkika, and Alessandra Fiorio Pla. TRPM8
inhibits endothelial cell migration via a non-channel function by
trapping the small GTPase Rap1. *Journal of Cell Biology*, 216
(7):2107–??, July 2017. CODEN JCLBA3. ISSN 0021-9525
org/content/216/7/2107.

Gong:2019:GCE


Gonzalez-Gaitan:2015:WCB


Ganguly:2017:HCA


Gilmore-Hall:2019:CPM

CCP1 promotes mitochondrial fusion and motility to prevent Purkinje cell neuron loss in pcd mice. Journal of Cell Biology, 218(1):206–??, January 2019. CODEN
REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/1/206.


REFERENCES


Gingras:2019:RBL


Glick:2017:NIP


Georgiadou:2017:ANR


Gao:2018:ART


Genna:2018:PFD

REFERENCES


REFERENCES


REFERENCES


REFERENCES

Gama:2017:MMD


Graef:2016:DMD


Gonzalez-Rodriguez:2019:XER


Gao:2018:NVA


Giacomello:2018:IOM


Guimaraes:2015:PLD

[GSB+15] Sofia C. Guimaraes, Martin Schuster, Ewa Bielska, Gulay Dagdas, Sreedhar Kilaru, Ben R. A. Meadows, Michael Schrader,


**Gopal:2015:TPC**


**Gagliardi:2018:MAC**


**Gomez-Sanchez:2018:AEA**


**Goo:2017:ADT**


Gowrishankar:2017:IJD


Gan:2019:AAT


Gu:2016:PTP


Xia:2015:DHC


Xia:2016:CDH

Hong guang Xia, Ayaz Najafov, Jiefei Geng, Lorena Galan-Acosta, Xuemei Han, Yuan Guo, Bing Shan, Yaoyang Zhang, Erik Norberg, Tao Zhang, Lifeng Pan, Junli Liu, Jonathan L. Coloff, Dmitry Ofengeim, Hong Zhu, Kejia Wu, Yu Cai, John R. Yates, Zhengjiang Zhu, Junying Yuan, and Helin
REFERENCES


**Gao:2018:VLT**


**Gorur:2017:CCM**


**Goult:2018:TMS**


**Haynes:2015:GCB**


**Hall:2015:CFY**

REFERENCES

Hammond:2018:DMI


Hayward:2019:CCC


Hyenne:2015:RCM


Hardin:2016:RCC


Hawkins:2018:DMC


Hu:2016:CAR

REFERENCES

Hutchins:2018:DAM

Howell:2015:WBP

Hayward:2019:CSE

Hendrix:2015:LCO
REFERENCES


Hau:2017:MHP


Hached:2019:EAD


Holtz:2015:SHI


Hofbauer:2018:MRP


Hos:2017:TIE

[HGG⁺¹⁷] Nina Judith Hos, Raja Ganesan, Saray Gutiérrez, Deniz Hos, Jennifer Klimek, Zeinab Abdullah, Martin Krönke, and Nirmal Robinson. Type I interferon enhances necroptosis of Salmonella


REFERENCES

Horton:2016:MFS

Hansen:2018:MAD

Heusermann:2016:ESF

Heald:2015:TYS

Hamdan:2017:SCW
Norfadilah Hamdan, Paraskevi Kritsiligkou, and Chris M. Grant. ER stress causes widespread protein aggregation and

Hagiwara:2018:CPC


Hamm:2016:SCC


Homma:2019:CKA


Hennig:2015:PLD

REFERENCES


REFERENCES


[HOH+16] Ari Hashimoto, Tsukasa Oikawa, Shigeru Hashimoto, Hirokazu Sugino, Ayumu Yoshihawa, Yutaro Otsuka, Haruka Handa, Yasuhito Onodera, Jin-Min Nam, Chitose Oneyama, Masato

Hanson:2019:PSP


Holzer:2019:RCI


Hong:2017:PCM


Hong:2015:PPC


Marie-Louise Hammarskjold and David Rekosh. SR proteins: To shuttle or not to shuttle, that is the question. Journal of Cell Biology, 216(7):1875–??, July 2017. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/7/1875.


REFERENCES


[HTLG18] Jennifer L. Hu, Michael E. Todhunter, Mark A. LaBarge, and Zev J. Gartner. Opportunities for organoids as new models
REFERENCES


REFERENCES


REFERENCES


REFERENCES


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/3/725.


REFERENCES

January 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/1/211.

Ishikawa:2017:UTB


Isokane:2016:AES


Yamashita:2016:MDO


Iuliano:2018:MPA

REFERENCES


REFERENCES

ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/9/2859.

[JBMM16] Alexis A. Jourdain, Erik Boehm, Kinsey Maundrell, and Jean-Claude Martinou. Mitochondrial RNA granules: Compartmen-


[JDG16] Iris K. Jarsch, Frederic Daste, and Jennifer L. Gallop. Mem-

[JDZ+16] Shawn N. Jordan, Tim Davies, Yelena Zhuravlev, Julien Du-
mont, Mimi Shirasu-Hiza, and Julie C. Canman. Cortical PAR polarity proteins promote robust cytokinesis during asymmet-
REFERENCES


Juanes:2019:RAM


Judith:2019:ASF


Jin:2017:EPS


Johnson:2015:FAB


Juettner:2019:VPS

Vanessa V. Juettner, Kevin Kruse, Arkaprava Dan, Vinh H. Vu, Yousaf Khan, Jonathan Le, Deborah Leckband, Yulia Komarova, and Asrar B. Malik. VE-PTP stabilizes VE-cadherin junctions and the endothelial barrier via a phosphatase-independent mechanism. *Journal of Cell Biology*, 218(5):1725–??, May 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/213/6/602.


REFERENCES

ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/214/1/2.


REFERENCES

DEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/212/1/51.


Karasu:2019:CBP


Kubo:2015:SCI


Klapproth:2019:KLP


Kasula:2016:MDH


Keeler:2017:EOA

REFERENCES

Kuhn:2017:SAC


Kavaler:2018:MSN


Knoops:2015:YPC


Kam:2018:DMG


Kendrick:2019:HSO

Kuhn:2015:RBI


Kelly:2016:HMM


Karg:2017:CKM


Keyes:2018:SCA


Konig:2017:MRD


Kim:2015:AFR

Nam-Gyun Kim and Barry M. Gumbiner. Adhesion to fibronectin regulates Hippo signaling via the FAK–Src–PI3K


[KHS+16] Sayaka Katsunuma, Hisao Honda, Tomoyasu Shinoda, Yuki-taka Ishimoto, Takaki Miyata, Hiroshi Kiyonari, Takaya Abe,


REFERENCES


Andrew Kodani, Tyler Moyer, Allen Chen, Andrew Holland, Christopher A. Walsh, and Jeremy F. Reiter. SF11 promotes centriole duplication by recruiting USP9X to stabilize the microcephaly protein STIL. *Journal of Cell Biology*, 218(7):2185–??, July 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/7/2185.


REFERENCES


[KPA+16] Nancy Kedersha, Marc D. Panas, Christopher A. Achorn, Shawn Lyons, Sarah Tisdale, Tyler Hickman, Marshall Thomas, Judy Lieberman, Gerald M. McInerney, Pavel Ivanov, and Paul


Kovacevic:2018:CRK


Kuri:2017:CDV


Kuri:2017:DVA


Kumar:2019:FMI


Kannan:2015:CSC

Nivetha Kannan and Vivian W. Tang. Correction: Synaptopodin couples epithelial contractility to α-actinin-4-dependent
REFERENCES


Min-Ji Kang, Deepika Vasudevan, Kwonyoon Kang, Kyunggon Kim, Jung-Eun Park, Nan Zhang, Xiaomei Zeng, Thomas A. Neubert, Michael T. Marr, and Hyung Don Ryoo. 4e-BP is a target of the GCN2–ATF4 pathway during *Drosophila* development and aging. *Journal of Cell Biology*, 216(1):115–??, Jan-
Klare:2015:CCB


Kim:2015:MAM


Kelliher:2018:AKE


Kim:2018:CSR


Lacefield:2019:DTR

REFERENCES


REFERENCES


[LDM15] Anne-Marie Ladouceur, Jonas F. Dorn, and Paul S. Maddox. Mitotic chromosome length scales in response to both cell and


REFERENCES

Leslie:2015:CDA

Leslie:2015:DTG

Leslie:2015:EEC

Leslie:2015:EKR

Leslie:2015:FSS

Leslie:2015:HIG

Leslie:2015:HCS

Mitch Leslie. Hsc70’s trip to the tip. *Journal of Cell Biology*, 210(5):??, August 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/210/5/682.3.


REFERENCES

Leslie:2015:NJF

Leslie:2015:NFS

Leslie:2015:NSI

Leslie:2015:PCC

Leslie:2015:PPA

Leslie:2015:PKF

Leslie:2015:SWG
REFERENCES

Leslie:2015:SRP

Leslie:2015:STE

Leslie:2015:SGE

Leslie:2015:SSF

Leslie:2015:TPC

Leslie:2015:TCD

Leslie:2015:TMU
REFERENCES


REFERENCES


REFERENCES

Lee:2016:GPP


Liu:2018:GDP


Lee:2018:RMN


Lamech:2015:UPA


Leandro:2019:SLD

REFERENCES


REFERENCES


REFERENCES

LeBlanc:2017:DSC


Liang:2018:ARE

Jin Rui Liang, Emily Lingeman, Saba Ahmed, and Jacob E. Corn. Atlastins remodel the endoplasmic reticulum for selective autophagy. *Journal of Cell Biology*, 217(10):3354–??, October 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/10/3354. See correction [LLAC18b].

Liang:2018:CAR


Longmate:2017:SIE


Lai:2017:HSE

Lin:2015:LRD


Liu:2018:UPK


Le:2016:THR


Lu:2018:OFC


Lim:2015:CPH


REFERENCES

Li:2019:HC

Lopes:2018:CAA

Lelli:2016:CIM

Lobato-Marquez:2016:SRM

Leyme:2015:IAT


REFERENCES


Lawson:2018:RGS


LeClaire:2015:NIK


Ladang:2015:EDW


Lakoduk:2019:MPA


Ladouceur:2017:CTI
REFERENCES

Liu:2016:TTR


Li:2018:ELD


Li:2015:NTA


Lee:2018:PUR


Lee:2018:BMW

REFERENCES


Lawrence:2016:LCP


Li:2018:CBC


Lee:2018:AFP


Lord:2015:ANP


REFERENCES

ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/215/1/5.


REFERENCES


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/214/6/653.


REFERENCES


Moyer:2015:BSP


Malinova:2017:AUS


McMurray:2019:LSM


Moriel-Carretero:2017:FAF


Meng:2015:RUS

Qingcai Meng, Chunmei Cai, Tingzhe Sun, Qianliang Wang, Weihong Xie, Rougfu Wang, and Jun Cui. Reversible ubiquitination shapes NLRC5 function and modulates NF-κB activation switch. *Journal of Cell Biology*, 211(5):1025–??, December


REFERENCES


REFERENCES

McHugh:2018:MET


Miller:2015:GNM


Magidson:2016:UKR


Miyagawa:2019:MMR


Matson:2019:ICD

Jacob Peter Matson, Amy M. House, Gavin D. Grant, Huaitong Wu, Joanna Perez, and Jeanette Gowen Cook. Intrinsic checkpoint deficiency during cell cycle re-entry from quiescence.
Maeshima:2018:CUM


Morita:2018:GWC


Mast:2018:EIR


Milev:2015:TTP


Miroshnikova:2019:CBM

Yekaterina A. Miroshnikova, Tim Hammesfahr, and Sara A. Wickström. Cell biology and mechanopathology of lamino-


Mario Mauthe, Martijn Langereis, Jennifer Jung, Xingdong Zhou, Alex Jones, Wienand Omta, Sharon A. Tooze, Björn Stork, Søren Riis Paludan, Tero Ahola, Dave Egan, Christian


REFERENCES


REFERENCES

Miteva:2019:RIC


Marcos-Ramiro:2016:RCE


Mostofa:2018:CCS


McNally:2018:MSE

REFERENCES


Manil-Segalen:2018:CSA


Moradi:2017:DRA


Murley:2015:LLS


Mattaini:2016:ISM


Mohan:2019:DMS

(print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/2/632.

Monachino:2017:WCM


Melloy:2007:NFD


Melloy:2017:CNF


Mozzetta:2019:CCH


Mizutani:2017:OIO

Ma:2019:CCG


Medeiros:2018:ABM


Mathew:2017:SAS


Ma:2016:CCN


McConnell:2016:RFE


Moudry:2016:TRR


Mathur:2017:MSB


Mason:2016:RGI


Murley:2017:STM


Mannen:2016:SNB


REFERENCES


Niewidok:2018:SMI


Niethammer:2016:NMG


Niedergang:2019:DCM


Nilsson:2019:PPR


Nakamura:2019:PRP

Nishimura:2016:DSE


Nakajo:2016:ECR


Nishimura:2019:GAR


Na:2015:TRC

REFERENCES


REFERENCES


[NOS+15] Satoshi Ninagawa, Tetsuya Okada, Yoshiki Sumitomo, Satoshi Horimoto, Takehiro Sugimoto, Tokiro Ishikawa, Shunichi Takeda, Takashi Yamamoto, Tadashi Suzuki, Yukiko Kamiya,


REFERENCES


[Nob19] Nathaniel Noblett, Zili Wu, Zhao Hua Ding, Seungmee Park, Tony Roenspies, Stephane Flibotte, Andrew D. Chisholm, Yishi Jin, and Antonio Colavita. DIP-2 suppresses ectopic neurite


ODonnell:2019:LFL

Olenick:2019:DAH

Ommer:2019:RGS

O'Connor-Giles:2016:TTT

ODonnell:2018:JEC
Ogawa:2018:NTA


Okatsu:2015:PUC


O'Neill:2016:UEE


Ohsaki:2016:PII


Ouenzar:2017:CCD

Oury:2019:MLR


Orr:2019:NCL


Oku:2017:EEC


Otera:2016:DDM


Otani:2019:CJC

REFERENCES


[OSL+19] Ellen C. O’Shaughnessy, Orrin J. Stone, Paul K. LaFosse, Mihai L. Azoitei, Denis Tsygankov, John M. Heddleston, Wes-

[ORegan:2015:HTM]


[OConnor:2017:DMF]


[Ogungbenro:2018:CCP]


[Ott:2016:MRL]

Carolyn M. Ott. Midbody remnant licenses primary cilia formation in epithelial cells. *Journal of Cell Biology*, 214(3):237–??, August 2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-
Osorio:2019:IRN


Pereira:2019:PRD


Pereira:2016:IFS


Paul:2015:IRP


Pinto:2016:PCP


REFERENCES


REFERENCES


Pedersen:2015:TRM


Panas:2019:PGS


Prosperi:2015:MFE


Pierobon:2017:UUF


Parisi:2015:ARM

Papadopoulous:2018:PTN


Park:2015:ANM


Prosser:2015:CRC


Pai:2018:DAD


Pontes:2017:MTC

REFERENCES


REFERENCES


Powell:2015:PST


Powell:2015:SHB


Powell:2015:TPS


Powell:2016:EDP


Powell:2016:EPC


Powell:2016:JNC


Vittoria Pagliarini, Laura Pelosi, Maria Blaire Bustamante, Annalisa Nobili, Maria Grazia Berardinelli, Marcello D'Amelio, Antonio Musaro, and Claudio Sette. SAM68 is a physiological regulator of SMN2 splicing in spinal muscular atrophy. *Journal of Cell Biology*, 211(1):77–??, October 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/211/1/77.


Emily J. Platt, Leslie Smith, and Mathew J. Thayer. L1 retrotransposon antisense RNA within ASAR lncRNAs controls chromosome-wide replication timing. *Journal of Cell Bi-

References
Pamula:2016:SCH

Platani:2015:MDL

Palmer:2019:CRN

Pugh:2015:PDM

Pacquelet:2015:PAR
Anne Pacquelet, Perrine Uhart, Jean-Pierre Tassan, and Grégoire Michaux. PAR-4 and anillin regulate myosin to coordinate spindle and furrow position during asymmetric division.
Potapova:2019:SMR

Perez-Vale:2018:MAB

Park:2019:PCG

Prasad:2018:HCA

Pina:2018:SAE


REFERENCES

Qu:2017:PMR


Qu:2017:SDM


Qi:2016:SHM


Qu:2019:SBS


Rabouille:2017:KCR

REFERENCES


Razzell:2018:FSP


Rosenberg:2015:MRH


Riccio:2019:DEU


Richter:2019:RCH


Roberson:2015:TMO


[RHH\+18] Léa Ripoll, Xavier Heiligenstein, Ilse Hurban, Lia Domingues, Florent Fignon, Karl J. Petersen, Megan K. Dennis, Anne Houdusse, Michael S. Marks, Graça Raposo, and Cédric Delevoye. Myosin VI and branched actin filaments mediate...
REFERENCES


**Ren:2018:DRS**


**Randrianarison-Huetz:2018:SCS**


**Rice:2018:NLG**


**Risher:2018:TRP**

REFERENCES


REFERENCES

Raote:2019:PTV

Resende:2018:AIS

Renault-Mihara:2017:RRS

Romero-Morales:2017:APL

Rhys:2018:LCP
<table>
<thead>
<tr>
<th>Reference</th>
<th>Authors</th>
<th>Title</th>
<th>Journal</th>
<th>Volume Issue Page</th>
<th>Date</th>
<th>URL</th>
</tr>
</thead>
<tbody>
<tr>
<td>RMTR17</td>
<td>Luisa Robbez-Masson, Christopher H. C. Tie, and Helen M. Rowe</td>
<td>Cancer cells, on your histone marks, get SETDB1, silence retrotransposons, and go!</td>
<td>Journal of Cell Biology</td>
<td>216(11) 3429-??</td>
<td>November 2017</td>
<td><a href="http://jcb.rupress.org/content/216/11/3429">link</a></td>
</tr>
<tr>
<td>RND+17</td>
<td>Diana Rüthnick, Annett Neuner, Franziska Dietrich, Daniel Kirrmaier, Ulrike Engel, Michael Knop, and Elmar Schiebel</td>
<td>Characterization of spindle pole body duplication reveals a regulatory role for nuclear pore complexes.</td>
<td>Journal of Cell Biology</td>
<td>216(8) 2425-??</td>
<td>August 2017</td>
<td><a href="http://jcb.rupress.org/content/216/8/2425">link</a></td>
</tr>
<tr>
<td>RNP+17</td>
<td>Julia D. Romano, Sabrina J. Nolan, Corey Porter, Karen Ehrenman, Eric J. Hartman, Ru ching Hsia, and Isabelle Coppens</td>
<td>The parasite Toxoplasma sequesters diverse Rab host vesicles within an intravacuolar network.</td>
<td>Journal of Cell Biology</td>
<td>216(12) 4235-??</td>
<td>December 2017</td>
<td><a href="http://jcb.rupress.org/content/216/12/4235">link</a></td>
</tr>
<tr>
<td>RO18</td>
<td>Pierre Romé and Hiroyuki Ohkura</td>
<td>A novel microtubule nucleation pathway for meiotic spindle assembly in oocytes.</td>
<td>Journal of Cell Biology</td>
<td>217(10) 3431-??</td>
<td>October 2018</td>
<td><a href="http://jcb.rupress.org/content/217/10/3431">link</a></td>
</tr>
<tr>
<td>Roy16</td>
<td>Subhojit Roy</td>
<td>Waves, rings, and trails: The scenic landscape of axonal actin.</td>
<td>Journal of Cell Biology</td>
<td>212(2) 131-??</td>
<td>January 2016</td>
<td><a href="http://jcb.rupress.org/content/212/2/131">link</a></td>
</tr>
</tbody>
</table>
| RPMC+16   | Ajay Ramesh, Valentina Peleh, Sonia Martinez-Caballero, Florian Wollweber, Frederik Sommer, Martin van der Laan,

[Riemer:2017:OCP]


[Raiborg:2016:PMR]


[Rickman:2019:AUD]


[Ratcliffe:2019:HIM]

Roarty:2015:RRB


Reyes:2015:ABP


Rutkowski:2015:CBF


Roelants:2015:PKG


Ren:2019:ECS


REFERENCES

3171–??, October 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/10/3171.


REFERENCES

Sarouim:2015:NBM

Sarouim:2015:NBM
Schepis:2018:PSR

Schepis:2018:PSR
Shrima:2017:DKM

Shrima:2017:DKM
Schwille:2015:JSL

Schwille:2015:JSL
Schlett:2017:MTM


REFERENCES

Senaratne:2017:AOD


Sharma:2019:CA


Simonetti:2017:SDC


Saez:2019:ECL


Strale:2015:FONa


REFERENCES


REFERENCES


[Sed15m] Caitlin Sedwick. Growing up and growing pores in myoblast fusion. Journal of Cell Biology, 211(1):??, October 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/211/1/2.3.


Caitlin Sedwick. mTORC2 tips the balance in cell survival. *Journal of Cell Biology*, 211(1):??, October 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/211/1/2.1.


Sedwick:2015:TSS


Sedwick:2016:JFM


Sedwick:2016:JKE


Sedwick:2016:MSS


Sedwick:2016:NBC


Sedwick:2016:SDU

References


[SES1+19] Nicole Scholz, Nadine Ehmann, Divya Sachidanandam, Cordelia Imig, Benjamin H. Cooper, Olaf Jahn, Kerstin Reim, Nils Brose, Jutta Meyer, Marius Lamberty, Steffen Altrichter, Anne

Scholz:2019:CCB

Schoenfelder:2015:EIP


Strothman:2019:MME


Schmidt:2017:TPC


Stroud:2017:NEM


Zhiqi Sun, Shengzhen S. Guo, and Reinhard Fässler. Integrin-mediated mechanotransduction. *Journal of Cell Biology*, 215...
REFERENCES


REFERENCES


REFERENCES


REFERENCES


Ben Short. NORE1A erects a senescence barrier to tumorigenesis. *Journal of Cell Biology*, 208(6):??, March 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/208/6/650.2.


REFERENCES


REFERENCES


REFERENCES


REFERENCES


[Sho16g] Ben Short. Decrypting a collagen’s role in schizophrenia. *Journal of Cell Biology*, 212(6):??, March 2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/212/6/606.3.


REFERENCES

353


[Sho16r] Ben Short. MLL5 limits PLK1 aggregation. *Journal of Cell Biology*, 212(7):??, March 2016. CODEN JCLBA3. ISSN
REFERENCES

Ben Short. NBR1 helps autophagosomes take a bite out of focal adhesions. *Journal of Cell Biology*, 212(5):??, February 2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/212/5/482.3.


REFERENCES


**Short:2016:RAS**


**Short:2016:RCP**


**Short:2016:SHS**


**Short:2016:SPB**


**Short:2016:SF**


**Short:2016:SSG**


**Short:2016:SSS**

REFERENCES


[Sho17b] Ben Short. Centrosome signaling pathways consult on their decision. *Journal of Cell Biology*, 216(9):2599–??, September


REFERENCES


Short:2018:PHE


Short:2018:SIR


Short:2018:STF


Sing:2018:BYR


Smith:2017:MCD


Spiess:2018:AII

Saunders:2017:TCT


Selyunin:2017:GWS


Shigematsu:2018:SIM


Siletti:2016:AML


Siletti:2016:RGP

Siletti:2017:RMM


Scharaw:2016:ETR


Sobajima:2018:RBP


Silva:2016:ULI


Sawyer:2019:DRO


REFERENCES

Shimi:2019:MOY


Smoyer:2016:AMP


Sidoli:2017:WPN


Singh:2018:SSM


Shima:2019:CVC

Son:2015:RMV


Saraya:2015:PRH


Seibert:2019:CMP


Sohn:2018:PPC


Song:2018:MHA


Song:2019:PTR


Sikorska:2016:LQC


Smith:2015:VSP


Slep:2016:MDR


Sun:2018:TIT

[SLG+18] Hao Sun, Frederic Lagarrigue, Alexandre R. Gingras, Zhichao Fan, Klaus Ley, and Mark H. Ginsberg. Transmission of integrin $\beta\gamma$ transmembrane domain topology enables gut lymphoid


[SM18] Leonid Serebryannyy and Tom Misteli. Protein sequestration at the nuclear periphery as a potential regulatory mechanism in premature aging. *Journal of Cell Biology*, 217(1):21–??, January 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-


REFERENCES

Silva:2016:RGR


Sorensen:2017:RWR


Simoes:2017:MIP


Salinas:2017:PDF


Schietroma:2017:UST

REFERENCES

June 2017. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/6/1849.


Christoph Schiklenk, Boryana Petrova, Marc Kschonsak, Markus Hassler, Carlo Klein, Toby J. Gibson, and Christian H. Haering. Control of mitotic chromosome condensation by the fission yeast transcription factor Zas1. *Journal of Cell Biology,*
REFERENCES


REFERENCES


REFERENCES


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/6/1659.


Schvartzman:2018:MRC

Sampayo:2018:FRE

Schroeder:2016:AAD

Schreiber:2015:ULT

Sargent:2016:PEU
REFERENCES


REFERENCES

3320–??, October 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/10/3320.


REFERENCES


Tardieux:2016:RMP


Tarasenko:2017:MCM


Tang:2016:SHF


Thangavel:2015:DDP


Tornavaca:2015:ZCE

Olga Tornavaca, Minghao Chia, Neil Dufton, Lourdes Osuna Almagro, Daniel E. Conway, Anna M. Randi, Martin A. Schwartz, Karl Matter, and Maria S. Balda. ZO-1 controls endothelial adherens junctions, cell–cell tension, angiogenesis,

**Terawaki:2015:RFD**


**Tang:2018:PIS**


**Thuma:2018:DIC**


**Trudeau:2016:LAP**


DEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/1/10.


REFERENCES


Vitor Teixeira, Lisa Johnsen, Fernando Martínez-Montañés, Alexandra Grippa, Laura Buxó, Fatima-Zahra Idrissi, Christopher S. Ejsing, and Pedro Carvalho. Regulation of lipid droplets


[TM18] Nicole M. Templeman and Coleen T. Murphy. Regulation of reproduction and longevity by nutrient-sensing pathways.
 REFERENCES


Tokarz:2018:CBS


Tsoumekos:2018:DBB


Tavernier:2015:CPS


Tooze:2018:SAR


Trainees:2018:GBP

REFERENCES


April 2015. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/209/1/33.

Tian:2015:ISH


Tsuchiya:2018:IXP


Tsuchiya:2019:CIX


Taniguchi:2017:AIS


Tourriere:2019:RPG

[TT19] Hélène Tourrière and Jamal Tazi. Reply to “Phosphorylation of G3BP1-S149 does not influence stress granule assem-


REFERENCES


REFERENCES


REFERENCES

vonBudingen:2015:MOG


Vandersmissen:2015:EMT


vanDrogen:2019:MSI


vanderVaart:2017:TSE

Verlhac:2016:MCK

Marie-Hélène Verlhac. Mother centrioles are kicked out so that starfish zygote can grow. Journal of Cell Biology, 212 (7):759–??, March 2016. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/212/7/759.

Verlhac:2018:ASD


Volkov:2015:CPF


Vos:2017:CPE


vanGisbergen:2018:ASF

REFERENCES


Vogtle:2015:FLP


Viero:2015:TDR


vanLeeuwen:2018:MSP


Vichas:2015:SFH


Verma:2019:DOB


Verlhac:2018:STM


VanderLugt:2017:TDT


Venditti:2019:MDG


vanSteensel:2015:SGT


Velazquez:2016:LDM

vanVugt:2017:SPS

Vijayan:2017:APC

Viotti:2018:SER

Vossel:2015:TRP

Venkei:2018:EMA
Valverde:2019:ATL


Victoria:2017:SPL


Vermunt:2019:IGR


Volin:2018:MSP


Warren:2015:POM


Wu:2018:AMC


[WB18]
REFERENCES

ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/10/3531.


Wang:2015:YRR

Webb:2017:GEP

Wenz:2015:SCF

Wynne:2015:KFC

Wakatsuki:2015:OSD
Shuji Wakatsuki, Akiko Furuno, Makiko Ohshima, and Toshiyuki Araki. Oxidative stress-dependent phosphorylation activates ZNRF1 to induce neuronal/axonal degeneration.
Westhorpe:2015:CFC


Wagner:2016:LRA


White:2018:CPS


Wissel:2018:TRT


Winsor:2017:CCS

REFERENCES

Walther:2018:QMH

Wang:2019:CCA

Wilkinson:2015:BCB

Wood:2017:CHR

Watanabe:2015:TER


REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/2/477.


REFERENCES


Wang:2018:IMT


Wang:2018:SMP


Wei:2017:SII


Wang:2018:ISL


Wan:2018:PPD

[WWC+18] Zhengpeng Wan, Chenguang Xu, Xiangjun Chen, Hengyi Xie, Zongyu Li, Jing Wang, Xingyu Ji, Haodong Chen, Qinghua Ji, Samina Shaheen, Yang Xu, Fei Wang, Zhuo Tang, Ji-Shen


REFERENCES

[XLW\textsuperscript{+18}] Dijin Xu, Yuqi Li, Lizhen Wu, Ying Li, Dongyu Zhao, Jinhai Yu, Tuozhi Huang, Charles Ferguson, Robert G. Parton, Hongyuan Yang, and Peng Li. Rab18 promotes lipid droplet (LD) growth by tethering the ER to LDs through SNARE and NRZ interactions. *Journal of Cell Biology*, 217(3):975–??, March 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/3/975.


[XRH\textsuperscript{+18a}] Wanqing Xiang, M. Julia Roberti, Jean-Karim Hériche, Sébastien Huet, Stephanie Alexander, and Jan Ellenberg. Correction: Correlative live and super-resolution imaging reveals the dynamic structure of replication domains. *Journal of Cell Biology*, 217(9):3315–??, September 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/217/9/3315. See [XRH\textsuperscript{+18b}].

[XRH\textsuperscript{+18b}] Wanqing Xiang, M. Julia Roberti, Jean-Karim Hériche, Sébastien Huet, Stephanie Alexander, and Jan Ellenberg. Cor-


[XWZ+15] Chuan-Ming Xie, Dongping Wei, Lili Zhao, Sylvie Marchetto, Lin Mei, Jean-Paul Borg, and Yi Sun. Erbin is a novel substrate of the Sag-βTrCP E3 ligase that regulates KrasG12D-induced
REFERENCES


Yanakiev a:2019:CTM


Yang:2017:DRF


Yao:2017:VEP


Yamada:2015:RCB


Yin:2017:GPA

[YHG+17] Jianhua Yin, Yaling Huang, Pengfei Guo, Siqi Hu, Sawako Yoshina, Nan Xuan, Qiwen Gan, Shohei Mitani, Chonglin Yang, and Xiaochen Wang. GOP-1 promotes apoptotic cell

**Yasunaga:2015:PPI**


**Yukawa:2015:MWP**


**Yu-Kemp:2017:CEE**


**Yin:2016:PPD**


Patricia L. Yeyati, Rachel Schiller, Girish Mali, Ioannis Kasioulis, Akane Kawamura, Ian R. Adams, Christopher Playfoot, Nick Gilbert, Veronica van Heyningen, Jimi Wills, Alex von Kriegsheim, Andrew Finch, Juro Sakai, Christopher J. Schofield, Ian J. Jackson, and Pleasantine Mill. KDM3A coordinates actin dynamics with intraflagellar transport to regulate


REFERENCES

2017. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/216/6/1557.


REFERENCES


Zobel:2018:NEA


Zhu:2016:MPC


Zhuo:2015:URK


Zhang:2019:KSL

Zhao:2019:PCA


Zhu:2017:WKP


Zhang:2017:BCD


Zhao:2015:MNS


zurLage:2018:CDM

[zLSSS+18] Petra zur Lage, Panagiota Stefanopoulou, Katarzyna Styczynska-Sozcka, Niall Quinn, Girish Mali, Alex von Kriegsheim, Pleasantine Mill, and Andrew P. Jarman. Ciliary dynein motor preassembly is regulated by Wdr92 in association with HSP90 co-chaperone, R2TP. *Journal of Cell Biology*, 217(7):2583–??, July 2018. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-
REFERENCES

8140 (electronic). URL http://jcb.rupress.org/content/217/7/2583.

Zhao:2016:MMS


Zhang:2018:BBL


Zhang:2015:FDR


Zatulovskiy:2015:MS


Zhou:2015:PFS


REFERENCES

Zhou:2019:LCS


Zhou:2019:RDA

Fan Zhou, Zulin Wu, Mengzhu Zhao, Rakhilya Murtazina, Juan Cai, Ao Zhang, Rui Li, Dan Sun, Wenjing Li, Lei Zhao, Quili Li, Jing Zhu, Xiaoxia Cong, Yiting Zhou, Zhiping Xie, Valeriya Gyurkovska, Liuju Li, Xiaoshuai Huang, Yanhong Xue, Liangyi Chen, Hui Xu, Haiqian Xu, Yongheng Liang, and Nava Segev. Rab5-dependent autophagosome closure by ESCRT. *Journal of Cell Biology*, 218(6):1908–??, June 2019. CODEN JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/218/6/1908.

Zhang:2016:DMA


Zhang:2017:CAM


Zhou:2016:FAR

REFERENCES

JCLBA3. ISSN 0021-9525 (print), 1540-8140 (electronic). URL http://jcb.rupress.org/content/214/1/103.


